PATENT SPECIFICATION

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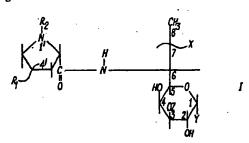
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(54) IMPROVEMENTS IN OR RELATING TO ANTIBIOTIC DERIVATIVES AND THE PREPARATION THEROF

(71) We, THE UPJOHN COMPANY, a corporation organized and existing under the laws of the State of Delaware, United States of America, of 301 Henrietta Street, Kalamazoo, State of Michigan, United States of America, do hereby declare the invention, for which we pray that a patent may be granted to us, and the method by which it is to be performed, to be particularly described in and by the following

This invention relates to novel antibacterial compounds and to processes for preparing them. It is particularly directed to novel 3-nucleotides of lincomycin, and of analogs thereof, and of celesticetins. The compounds of the invention can be shown by the following formula:



wherein Y can be in α or β configuration and is -SR wherein R is alkyl or 1 to 6 carbon atoms, inclusive,

or —S—CH₂—OH, R₁ is H, or cis or trans alkyl of from 1 to 8 carbon atoms, inclusive; R₂ is H, CH₃, or C₂H₃; X is OH, chlorine, bromine, iodine or —OR₃ wherein R₃ is alkyl of 1 to 6 carbon atoms, inclusive, cycloalkyl, hydroxyalkyl or alkoxyalkyl, each in the (R) or (S) configuration; and Z is a nucleoside -5'- phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine; and salts thereof. The invention also includes the zwitterion forms of compound I.

Alkyl of from 1 to 8 carbon atoms are methyl, ethyl, propyl, butyl, pentyl, hexyl, heptyl, and octyl and isomers thereof.

[Price 25p]

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ii.

5	The novel compounds of the invention are prepared by incorporating a compound, as defined in Formula I, wherein Z at the 3-position of the molecule is hydrogen (hereinafter referred to as the "parent" compound), in a Streptomyces fermentation, and transforming the compound into a novel 3-nucleotide, as described above. Also produced in varying amounts in the process of the subject invention, are 3-phosphate esters of the parent compounds. These 3-phosphate esters are readily distinguished from the 3-nucleotides of the subject invention since the 3-phosphates do not have an ultraviolet absorption maximum and they are not hydrolyzed to the parent compound by	. 5
10	snake venom diesterase (venom phosphodiesterase) or spicen diesterase. Thus, as which be described in detail hereinafter, the 3-nucleotides are detected in the recovery procedures of the subject invention by ultraviolet analysis and the snake venom diesterase test. Venom diesterase cleaves, for example, clindamycin-adenylate, to clindamycin and adecessing 50 phosphate.	
15	Initial presence of 3-nucleotides in fermentation beers is detected by the use of an alkaline phosphatase test, as hereinafter described. However, this test does not differentiate between 3-phosphates and 3-nucleotides and the 3-nucleotides would remain unrecognized but for the use of other tests, as described above. The companyed of the invention, though antibacterially inactive in vitro against	15
20	S. aureus and Sarcina lutea, are activated when used in vivo, for example against S. aureus. Presumably, this activation in vivo is comparable to the generation of the parent lincomycin compound when contacting the 3-nucleotide-lincomycin compound with alkaline phospharase in vitro. The lincomycin compounds, herein defined as starting materials or parent com-	20
25	pounds, can be prepared by procedures disclosed in various patents, publications and patent applications. These are as follows:	25
	Lincomycin With reference to Formula I, wherein Y = —SCH, to —SC ₄ H ₁ ; R ₁ =cis or trans alkyl to 8 carbon atoms U.S. Patent 3,086,912 U.S. Patent 3,380,992 U.S. Patent 3,380,992	
30	R ₂ =Hydrogen or alkyl to 8 carbon atoms X = (S)OH or OR ₂ X = (R) or (S) Cl or Br U.S. Paterx 3,380,992 U.S. Paterx 3,380,992 U.S. Paterx 676,202 U.S. Paterx 3,496,163 U.S. Paterx 3,496,163 U.S. Paterx 3,496,163	30
35	Celesticetin U.S. Patent 2,928,844 Desalicetin U.S. Patent 2,851,463	35
40	4'-Depropyl-4'-ethyl lincomycin, wherein Y is —SCH ₃ , R ₁ is trans ethyl, R ₂ is CH ₃ , and X is (R)OH in Formula I can be prepared by the procedure disclosed in Examples 1 and 2 of U.S. Patent 3,359,164 wherein said compound is named lincomycin B. 1'-Demethyl-1'-ethyl lincomycin, wherein Y is —SCH ₃ , R ₁ is trans n-propyl, R ₂ is ethyl, and X is (R)OH in Formula I can be prepared by the procedure disclosed in Examples 1 and 2 of U.S. Patent 3,359,163 wherein said compound is named linco-	40
45	mycin C. 1'-Demethyl lincomycin, wherein Y is —SCH ₂ , R ₁ is trans n-propyl, R ₂ is H and X is (R)OH in Formula I can be prepared by the procedure disclosed in Example 1 of U.S. Patent 3,329,568 wherein said compound is named lincomycin D. Of the above compounds, the compound 7(S)-chloro-7-deoxy-lincomycin is also presently known by the generic name "clindamycin".	45
50	The parent lincomycin compounds or analogs thereof, and celesticetin, as described above, can be converted to 3-nucleotides, as shown in Formula I, by incorporating the parent compound in a Streptomyces fermentation. For example, upon adding clindamycin hydrochloride to a Streptomyces coelicolor Müller, NRRL 3532, fermentation there are produced clindamycin nucleotides.	50
55	The fermentation to make the novel compounds of the invention can be conducted in an aqueous nutrient medium under submerged aerobic conditions. It is to be understood also that for the preparation of limited amounts of 3-nucleotides, surface cultures and bottles can be employed. The organism used in the fermentation is grown in a nutrient medium containing a carbon source, for example, an assimilable carbohydrate	55
60	and a nitrogen source, for example, an assimilable nitrogen compound or proteinaceous material. Preferred carbon sources include glucose, brown sugar, sucrose, glycerol, starch, comstarch, lactose, dextrin and molasses. Preferred nitrogen sources include corn steep liquor, yeast, autolyzed brewer's yeast with milk solids, soybean meal, cotton-	60

seed meal, commeal, milk solids, pancreatic digest of casein, distillers' solubles, animal peptone liquors and meat and bone scraps. Combinations of these carbons and nitrogen sources can be used advantageously. Trace metals, for example, zinc, magnesium, manganese, cubalt and iron need not be added to the fermentation media since tap water and unpurified ingredients are used as media components. 5 Production of the novel compounds of the invention can be effected at any temperature conducive to satisfactory growth of the Streptomyces culture, for example, between about 18° and 40°C., and preferably between about 20° and 37°C. When a Streptomycer fermentation, as described above, is used to prepare nucleo-tides of lincomycin or an analog thereof, as herein defined, or of celesticetin, the linco-10 10 mycin or celesticetin parent compound (non-nucleotide) can be added prior to inoculation of the fermentation medium. Alternatively, the parent compound can be added in small increments during the fermentation cycle so long as the addition is not too late in the fermentation cycle to accomplish the desired transformation of all the parent compound added. The time and amounts of addition of the parent compound 15 15 can easily be determined for each fermentation by adding the parent compound until some toxicity to the fermentation is observed, such as inhibition of the formation of 3-nucleotides. Also, if at the end of the fermentation cycle there remains parent compound, then in subsequent fermentations smaller levels of parent compound should 20 be used and/or the time of addition should be altered. 20 Since the in vitro antibacterial activity against S. Lutea of the parent compound is lost upon transformation to a 3-nucleotide, the presence of residual in vitro anti-bacterial activity in a culture or culture extract at 24 hours after addition of the parent compound is evidence that the capacity of the culture or culture extract to transform the parent compound has been exceeded or the level of added compound was too 25 25 high and inhibited the microorganism in the transformation process. The in vitro antibacterial activity, mentioned above, can be ascertained on a standard microbiological plate assay against the microorganism Sarcina lutea. A variety of procedures can be employed in the isolation and purification of the novel compounds in the subject invention, for example, solvent extraction, liquid-30 30 liquid distribution in a Craig apparatus, liquid ion exchange extraction or adsorption on a suitable adsorbent, for example, carbon, and column chromatography. In a preferred recovery process, the novel 3-nucleotide compounds are isolated from a fermentation beer, as herein described, by filtration. The filtrate is then passed over a suitable absorbent, for example, activated carbon or "Amberlite" (Registered Trade 35 35 Mark) XAD-2 (a non-ionic, macro-porous copolymer of styrene cross-linked with divinylbenzene resin sold by Rohm and Haas Company). This resin is prepared by suspension polymerization of styrene divinylbenzene copolymers in the presence of a substance which is a good solvent for the copolymer (see J.A.C.S. 84, 306, 1962) to remove water-soluble impurities which may interfere with the subsequent chromato-40 graphy step. The resin is eluted with a mixture of water and water-miscible organic graphy step. The resan is cluted with a mixture to water aim water-instrible organic solvents, for example, water-lower alcohols of C_1 — C_2 and water-lower ketones of C_3 — C_4 and water-lower ketones of C_4 — C_5 is then passed through a chromatography column containing an anion exchange resin, for example, "Dower"—1 (Registered Trade Mark) (X—4) in the acetate form (sold by Dow Chemical Company, Middland, Michigan). Fractions are collected from 45 45 the chromatography column and assayed for activity against the microorganism S. lutea before and after treatment of the fractions with alkaline phosphatase as hereinafter described. Fractions having the highest activity against S. lutea upon test with alkaline phosphatase are pooled, concentrated, then subjected to countercurrent distribution in 50 50 a Craig apparatus using a solvent system consisting of n-butanol-water (1:1 v/v). Fractions showing maximum ultraviolet absorption, and which are hydrolyzed by snake venom phosphodiesterase, are collected to give a preparation containing a mixture of 3-nucleotides. This mixture can be subjected to separation procedures to recover the individual 3-nucleotides. 55 55

A preferred separation procedure to recover the individual 3-nucleotides from a mixture thereof utilizes DEAE—"Sephadex"—Registered Trade Mark—(Pharmacia Fine Chemicals, Inc., Piscataway, N.J., U.S.A. or Pharmacia, Uppsala, Sweden) column chromatography. The column is eluted with tris-(hydroxymethyl)-amino-methane (THAM) acetate. Fractions are analyzed by testing for activity against S. lutea before and after alkaline phosphatase treatment and by ultraviolet spectrum analysis at the original pH of the fraction, and at an acid pH (ca. 2.0). Pools of fractions having biological activity against S. lutea after alkaline phosphatase treatment, and showing ultraviolet spectrum absorption, are made. Each pool contains a single 3-nucleotide,

along with other undesired materials. The THAM-acctate buffer is removed from these pools by passing them over a column containing "Amberlite" (Registered Trade Mark) XAD—2 packed in water. After washing the column with water, it is eluted with aqueous methanol (ca. 80% aqueous methanol). Fractions, about 20 ml. each, are collected and analyzed by U.V. Fractions showing U.V. absorption are combined and 5 concentrated to dryness to a residue. The residue is dissolved in a lower alcohol, for example methanol, and the solution mixed with ether to yield a precipitate of a 3-nucleotide compound, as defined in Formula I. As shown in Formula I, the 3-nucleotide moiety of the novel compounds of the invention are the 3-(5'-cytidylate), 3-(5'adenylate) 3-(5'-uridylate) and 3-(5'-guanylate).

The nucleotides can also be separated by partition chromatography over "Dicalite"

—Registered Trade Mark—(diatomaceous earth) using solvent systems consisting of 10 10 water and a water-immiscible solvent. Lincomycin 3-nucleotides and the 3-nucleotides of lincomycin analogues are essentially inactive against bacteria in vitro. Thus, these novel 3-nucleotide compounds are detected by testing for bioactivity against S. lutea after treatment of the samples 15 15 with alkaline phosphatase. For example, the reaction mixture consists of 0.5 ml. Tris buffer (0.5 M) pH 8.0, 0.5 ml. alkaline phosphatase (1 mg./ml.) stock made up in Tris buffer (0.5 M) pH 8.0, 0.05 ml. (about 50 mcg.) of lincomycin-3-nucleotides. This reaction mixture is incubated overnight at 28°C. 20 Illustrative of Streptomyces which can be used to prepare the novel compounds of the invention are S. coelicolor 1945, NRRL 3531; S. coelicolor Müller, NRRL 3532; and S. venezuelae, NRRL 3527. These cultures are available, without restriction, from the Northern Utilization and Research Division, Agricultural Research Service, U.S. Department of Agriculture, Peoria, Illinois, U.S.A. 25 The novel compounds of the invention are amphoteric compounds and can exist different ionic forms according to the pH of the environment. At low pH the compounds exist in the acid-addition salt form, at a higher pH in a zwitterion form, and at a still higher pH in a metal salt form. The acid-addition salts include those of strong organic or inorganic acids having a pK equal to or less than that of phosphate, 30 30 for example, hydrochloric, sulfuric and phosphoric acids.

Acid and metal salts include the alkali metal (e.g. Na and K), alkaline earth metal (e.g. Ca and Mg), Zn, Al and ammonium salts obtained by neutralizing an acid form with the appropriate base, for example, ammonium hydroxide, sodium and potassium hydroxides, or alkoxides, calcium, or magnesium hydroxides. The acid and neutral 35 35 salts also include amine salts obtained by neutralizing an acid form with a basic amine, for example, mono-, di-, and trimethylamines, mono-, di-, and triethylamines, mono-, di-, and tripropylamines (iso- and normal), ethyldimethylamine, benzyldiethylamine, cyclohexylamine, benzylamine, dibenzylamine, N,N'-dibenzylethylenediamine, bis-(ortho-methoxyphenylisopropyl)amine, and other lower-aliphatic, lower-cycloaliphatic, 40 and araliphatic amines, the lower-aliphatic and lower-cycloaliphatic radicals containing up to 8 carbon atoms; heterocyclic amines such as piperidine, morpholine, pyrrolidine, piperazine and the alkyl derivatives wherein the alkyl groups contain 1 to 8 carbon atoms, thereof such as 1-methylpiperidine, 4-ethylmorpholine, 1-isopropylpyrrolidine, 1,4-dimethylpiperazine, 1-n-butylpiperidine, 2-methylpiperidine and 1-ethyl-2-methyl-45 piperidine; amines containing water solubilizing or hydrophilic groups such as mono-, di-, and triethanolamines, ethyldiethanolamine, n-butyl monoethanolamine, 2-amino-1butanol, 2-amino-2-ethyl-1,3-propanediol, 2-amino-2-methyl-1-propanol, tris-(hydroxymethyl)-aminomethane, phenylmonoethanolamine, p-tertiaryamylphenyldiethanolamine, and galactamine, N-methylglucamine, N-methylglucosamine, ephedrine, phenylephrine, 50 epinephrine, and procaine; tetraethylammonium hydroxide; and guanidine. The various forms can be used interchangeably but for most purposes the zwitterion form

wherein R₁, R₂, X, Y and Z are as defined previously, and the ammonium salt form are preferred.

Preferably according to the invention there are provided compounds having the general formula:

and the saits thereof, wherein Y, R₁ and R₂ are as defined above and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine The invention also provides compounds having the general formula:

and the salts thereof wherein halo is chlorine or bromine and Y, R1, R2 and Z are as defined above.

Further the invention provides preferred compounds having the general formula:

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and salts thereof, wherein halo is chlorine or bromine, R, is CH₃; R₁ is pentyl; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine. Preferably in these latter compounds halo is chlorine.

Also provided by the present invention are compounds having the general formula:

wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine; R, is CH₂ and R₂ is pentyl.

Further preferred compounds of the present invention are those having the general

formula:

and salts thereof, wherein halo is chlorine or bromine; R₃ is CH₃; R₁ is pentyl; R₂ is hydrogen and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine,

guanosine, cytidine or uridine.

Further, the invention relates to a process for the therapeutic treatment of animals excluding humans hosting susceptible microbial disease-producing organisms (bacterial and other microparasites) and the prophylactic treatment of a disease-susceptible host comprising the administration of the 3-nucleotide esters or a pharmacologically acceptable salt to the host.

The compounds of the present invention are useful in the same manner as linco-mycin and celesticetin in the treatment of humans, birds and animals for various

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pathological conditions. The compounds provide a means for administering the therapeutic ingredient by the oral and parenteral routes for systemic treatment. The compounds provide a method of therapy for tonsillitis, pneumonia, otitis, conjunctivitis, boils, carbuncles and other infectious conditions of humans due to the presence of 5 bacteria. In animals, the compositions can be used prophylactically. For example, rats can be protected from Streptococcus viridans during shipment. Animals raised for 5 meat can be given prophylactic treatment for increased weight gain. Mammals hosting a parasitic protozoan of the class Sporazoa, order Coccidia (a microparasite producing the disease coccidiosis) can be treated by administration of the compounds of the present invention. For example cattle infected with E. zurnii, E. 10 10 bovis, E. illipsordalis; sheep and goats with E. parta, E. faurei; swine with E. debliecki, E. scabra, and Isospora siac; dogs and cans with Isospora bigemina, Isospora felis, E. canis, E. felini; poultry with E. tenella; rabbits with E. steedae, E. perforans; and mink with E. mustelae can be treated. The compounds are also useful in the treatment of diseases caused by members of 15 15 the genus Mycoplasma, the most commonly known forms are PPLO (pleuropneumonialike organisms) such as M. hominis, M. salivarium, M. mycoides, M. hyopneumonia, M. hyorhinis, M. gallisepticum, M. arthriditis and other species in man and animals, including domestic animals such as sheep, dogs, caule, swine, and poultry (e.g., chickens, turkeys, ducks, and geese) and laboratory animals (e.g., rats and mice). 20 The compositions find application in the treatment of kidney and other infections 20 when L forms of gram-negative and gram-positive bacteria are present, for example, L forms of P. mirabilis. 25 25 granules, pills, sterile parenteral solutions and suspensions, and oral solutions and suspensions, and oil-water emulsions. Powders are prepared by comminuting the 3-nucleotides to a suitably fine size and mixing with a similarly comminuted diluent. The diluent can be edible carbohydrate material such as starch or lactose. Advantageously, a sweetening agent or sugar 30 is present as well as a flavoring material. Dry granulations for reconstitution with water are prepared utilizing water-soluble diluents. A powder mixture of a finely divided 3-nucleotide and a water-soluble diluent such as sucrose or glucose, is wetted with a 30 binder such as acacia mucilage or gelatin solution and forced through a screen to form granules which are allowed to dry. Advantageously, a thickening or suspending agent 35 such as methylcellulose is present as well as a wetting agent and flavoring oil. 35 Capsules are produced by preparing a powder mixture as hereinbefore described and filling into formed gelatin sheaths. Advantageously, as an adjuvant to the filling operation, a lubricant such as tale, magnesium stearate and calcium stearate is added 40 to the powder mixture before the filling operation. Tablets are made by preparing a powder mixture, wet granulating or dry granulating or slugging, adding a lubricant, and pressing into tablets. The powder mixture 40 is prepared by mixing the 3-nucleotide suitably comminuted, with a diluent or base such as starch, lactose, kaolin or dicalcium phosphate. The powder mixture can be granulated by wetting with a binder such as corn syrup, gelatin solution, methylcellulose 45 solution or acacia mucilage and forcing through a screen. As an alternative granulating 45 procedure, the powder mixture can be slugged, i.e., run through the tablet machine procedure, the powder mixture can be singged, i.e., run through the tablet machine and the resulting large tablets (slugs) broken into granules. The granules can be lubricated to prevent sticking to the tablet-forming dies by means of the addition of stearic acid, a stearate sale, tale, or mineral oil. The lubricated mixture is then compressed 50 into tablets. Advantageously, the tablet can be provided with a protective coating consisting of a sealing coat of shellac, a coating of sugar and methylcellulose, and a polish coating of carnauba wax. Oral fluids are prepared in unit dosage forms such as syrups and elixirs wherein each teaspoonful of composition contains a predetermined amount of the 3-nucleotide 55 55 for administration. A syrup is prepared by dispersing the 3-nucleotide in a suitable flavored aqueous sucrose solution. Similarly, an clixir is prepared utilizing a hydro-alcoholic vehicle. Elixirs are advantageous vehicles for use when a solution is desired of a compound showing low solubility in water and good solubility in an aqueous-alcoholic medium. For parenteral administration, sterile fluid unit dosage forms can be prepared. In 60 preparing the parenteral form, a measured amount of the 3-nucleotide is placed in a vial; the vial and its contents sterilized and sealed. An accompanying vial of sterile water can be conveniently provided as a vehicle to form a suspension or solution

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(depending on water-solubility of compound) prior to administration. Advantageously the sterile water can have dissolved therein a suspending agent, local anesthetic, and buffering agents.

Alternatively, a parenteral suspension having prolonged action can be prepared by suspending the 3-nucleotide in a parenterally acceptable vegetable oil with or without

additional adjuvants.

The term "unit dosage form" as used in the specification and claims refers to physically discrete units suitable as unitary dosages for human subjects, each unit containing a predetermined quantity of active material calculated to the desired dosage in association with the required pharmaceutical diluent, carrier, or vehicle, such as e.g. a tablet, vial or ampoule. The specifications for the novel unit dosage forms of this invention are dictated by and are directly dependent on (a) the unique characteristics of the active material and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active material for therapeutic use as disclosed in detail in this specification, these being features of the present invention. Examples of suitable unit dosage forms in accord with this invention are tablets, capsules, powder packets, granules, wafers, ampuls, vials, segregated multiples of any of the foregoing, and other forms as herein described. The unit dosage forms comounded with a suitable pharmaceutical carrier contain, in the preferred embodiments, from 25 mg. to 500 mg. of 3-nucleotide or its pharmacologically acceptable salts per dosage unit and 5 to 65% w/v for parenteral preparations.

The amount of 3-nucleotide or salts thereof that is to be administered depends

on the age and weight of the patient, the particular condition to be treated, and the route of administration. A dose of from 1 mg./kg./day to 50 mg./kg./day is preferred

for systemic treatment.

Thus according to the invention there is provided a therapeutic composition comprising from 5% to 82% by weight of a compound of the general formula:

wherein R₁, R₂, Z, X and Y are as defined above or a pharmacologically acceptable salt thereof as an essential active ingredient in combination with a pharmaceutical vehicle.

The following Examples are illustrative of the process and products of the present invention, but are not to be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

Example 1 Clindamycin-3-Nucleotides

Fermentation

A soil stock of Streptomyces coelicolor Müller, NRRL 3532, is used to inoculate a series of 500-ml. Erlenmeyer flasks, each containing 100 ml. of sterile seed medium consisting of the following ingredients:

> 25 g./liter Glucose monohydrate 25 g./liter Balance Pharmamedia* Tap water q.s.
>
> *Pharmamedia is an industrial grade of cottonseed flour produced by Trader's Oil Mili
> Company, Fort Worth, Texas.

The shake flasks are grown for 3 days at 28°C. on a rotary shaker. Seed inoculum (5 ml.), prepared as described above, is used to inoculate each of

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	a series of 500-ml. Erlenmeyer flasks each containing 100 ml. of sterile fermentation medium consisting of the following ingredients:	
	Yeast Extract 2.5 g./liter	
	NZ amine B* 5.0 g./liter	
5 .	Glucose monohydrate 20 g./liter	5
	Sodium nitrate 3 g./liter	
	Dipotassium phosphate 1 g./liter Magnesium sulphate 0.5 g./liter	
	Magnesium sulphate 0.5 g./liser	
10	Potassium chloride 0.5 g./liter Ferrous sulphate 0.01g./liter	10
20	Tap water Balance	10
	*Sold by Difco Laboratories, Detroit, Michi-	
	gan. It is a bulk peptone in powder form	
	obtained by the pancreatic digestion of casein.	
15	100 mg/liter clindamycin hydrochloride is added to the fermentation flask broth	15
	24 hours after inoculation.	
	The fermentation flasks are grown for 24 hours at 28°C. on a rotary shaker. The transformation reaction in the fermentation flask is followed by measuring the loss of	
	clindamycin activity using an S. lutea standard curve assay. Approximately 100%	
20	of the added clindsmycin is transformed to an <i>in vitro</i> antibacterially inactive form in	20
	about 24 hours. The S. lutea assay is conducted as follows: The assay is on ager	20
	buffered to pH 6—8 with pH 7.0 phosphate buffer (0.1 M). A unit volume (0.08 ml)	
	of solution containing the material to be assayed is placed on a 12.7 mm. assay disc	
25	which is then placed on an agar plate seeded with the assay microorganism. The tray is incubated at 37°C. for 18—24 hours. In vitro antibacterial activity is evidenced	
	by 8 zone of growth inhibition surrounding the disc. Antihacterial activity can be	25
	expressed quantitatively as mov. parent compound (or as lincompound or clindomycin)/	
	mil by the linear relation of log dose to zone diameter, referred to the standard	
20	according to the art. Presence of clindsmycin-3-mideorides is determined by first	
30	incubating the inactive beer with alkaline phosphatase at pH 8.0 in Tris buffer, and	50
	then assaying the reaction mixture against S. lutea, as described above. B. Recovery	
	1) Filtration and Absorption on Non-Ionic Resin	
	The above fermentation is scaled up into a fermentation rank to produce 400 lives	
35	of fermentation beer containing clindamycin-3-nucleotides. The clindamycin-3-nucleo-	35
	tides are recovered from the whole beer by first filtering the whole beer with the	
	aid of 10 Kg. diatomaceous filter aid. The filter cake is washed with water. The aqueous wash is combined with the clear beer and the combined clear beer-wash is	
	treated with an adsorbent, for example carbon or Amberlite AXII	
40	and mass Company), in order to remove water-soluble improvings which send to and an analysis	40
	the efficiency of subsequent chromatography. The absorption column is personal by	40
	SULTVINE ADOUG 42 N.E. OF ADSOTDERY (Amberlite-Registered Tenda Made VATA 2)	
	in water, pouring the slurry into a glass column (2" inside diameter), allowing the slurry to settle under atmospheric pressure, and draining. The clear beer-wash, described	
45	above, is passed unrough the column at a now rate of shout 1 liter per minute. The	45
	column is washed with water: 100 liters of the water wash is discorded. The column is	40
	then clutted with 120 liters of 60% agreeus methanol (Ringra T) and 100 lines of	
	93% aqueous methanoi (Eruste 11). Eluste 1 is treated further to recover clindamycin-	
50	3-nucleotides, whereas Eluate II is discarded. 2) Absorption on Ion Exchange Resin	
•	Eluate I, described above, is chromatographed over an anion exchange above	50
	Engine Column. The column is mich with // Ka "Comer" Decision / T., J. 14 1	
	-(A-4) in the acetate form, supplied by Dow Chemical Company Middend Mid-	
55	Educate I is addusted to a Dri of 10.0 with concentrated ammonium budidd	
23	the alkaline solution is passed intollen the chromatography coheren. The grant limits	55
	from the column is concentrated to dryness; yield, 877 grams of material containing clindamycin-3-nucleotides. This material is labeled "Material A". The column is	
	uncil washed with 100 liters of water and chifed with 70 liters of 50/ company	
	duly life avoil avoi cluates are concentrated and the contrine concentrate is for an	
60	uried; yield 89.4 grams of material containing clindamycin-3-nucleotides. This material	60
•	is labeled "Material B".	

in 1.5 liters of water. The pH of the solution is adjusted to 7.3 with contentiated ammonium hydroxide and this solution is passed over a column containing 2 liters of "Amberlite" (Registered Trade Mark) XAD—2 liter fractions (W—1, W—2, W—3). The column is then eluced with 90% aqueous methanol. Fractions of 20 ml. are collected and tested for activity against 5. luteo before and after treatment with alkaline phosphanase. Fractions numbered 61—250 are combined and concentrated to dryness; yield, 52 g. of material containing clindamycin-3-nucleotides. This material is labeled "ADA—10.1". Fractions W—2 and W—3, described above, and fractions numbered 1—60 from the above Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—2 column, are combined and passed again over the same "Amberdite XAD—1 column, which is first regenerated with 15 liters of water (W—1 obtained as described above) and then eluted with 5 liters of absolute methanol. Three cuts are made, i.e. methanol fraction 1=1 liter; methanol fraction 3 = 3 liters. These fractions are tested for activity against S. lutea before and after treatment with alkaline phosphatase. Methanol fraction 3 is concentrated to dryness; yield, 12.63 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.2". Preparations ADA—10.1, ADA—11.1, and ADA—11.2 all prepared as described above, are combined as preparation ADA—37.1, described above, is dissolved in 30 lon ml. of up oper and 100 ml. of the lower phase of a solvent system consisting of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase ar		C. Purification 1) Absorption on Non-Ionic Resin	
liters of water. The aqueous wash is collected in three 2-liter riactions (w=1, w=2, w=3). The column is then cluted with 90%, aqueous methanol. Fractions of 20 ml. are collected and tested for activity against 5. Iuteo before and after treatment with alkaline phosphatuse. Fractions numbered 61—250 are combined and concentrated to dryness; yield, 52 g. of material containing clindamycin-3-nucleotides. This material is labeled "ADA—10.1". Fractions W—2 and W—3, described above, and fractions numbered 1—60 from the above Amberlite KAD—2 column, are combined and passed again over the same "Amberlite "(Registered Trade Mark) KAD—2 column, which is first regenerated with 5 liters of aver (W—1 obtained as described above) and then eluted with 5 liters of absolute methanol. Three curs are made, i.e. methanol fraction 1 =1 liter; methanol fraction 2 =1 liter; and, methanol fraction 3=3 liters. These fractions are tested for activity against S. Iutea before and after treatment with alkaline phosphatuse, Methanol fraction 2 is concentrated to dryness; yield, 12.63 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.1". Methanol fraction 3 is concentrated to dryness; yield, 12.63 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—12". Preparations ADA—3.1., ADA—11.1, and ADA—11.2, all prepared as described above, are combined as preparation ADA—37.1, (64.7 g.). This preparation containing clindamycin-3-nucleotides is purified further by counter double current distribution as described below. 2) Counter Double Current Distribution A portion (21 g.) of preparation ADA—37.1, described above, is dissolved in abuse, are collected for fractions. A total of 100 transfers are run. The collected fractions and the material in the CDCD mbes are analyzed for S. lutes activity before and after treatment with alkaline phosphatuse. Using the same conditions as above, two additional CDCD distributions are run, each of the above three distributions are remaining	5	commenters by describe and this solution is passed over a column containing 2 incres of	5
Fractions W—2 and W—3, described above, and fractions numbered 1—60 from the above Amberlite "(Registered Trade Mark) XAD—2 column, are combined and passed again over the same "Amberlite "(Registered Trade Mark) XAD—2 column, which is first regenerated with 15 liters of water (W—1 obtained as described above) and then cluted with 5 liters of absolute methanol. Three cuts are made, i.e. methanol fraction 1 = 1 liter; methanol fraction 2 = 1 liter; and, methanol fraction 3 = 3 liters. These fractions are tested for activity against S. lutea before and after treatment with alkaline phosphatase. Methanol fraction 2 is concentrated to dryness; yield, 12.63 g, of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.7". Methanol fraction 3 is concentrated to dryness; yield, 0.7 g, of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.2". Preparations ADA—10.1, ADA—11.1, and ADA—11.2, all prepared as described above, are combined as preparation ADA—37.1, (described above, is dissolved in a described below. 2) Counter Double Current Distribution A portion (21 g,) of preparation ADA—37.1, described above, is dissolved in 100 nl. of tup lower phase of a solvent system consisting of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase are collected in 50 ml. fractions. A total of 100 transfers are run. The collected fractions and the material in the CDCD ubes are analyzed for S. lutea activity before and after treatment with alkaline phosphatase. Using the same conditions as above, two additional CDCD distributions are run, each using 21 g of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: 10	10	liters of water. The aqueous wash is collected in three 2-liter fractions (W—1, W—2, W—3). The column is then eluted with 90% aqueous methanol. Fractions of 20 ml. are collected and tested for activity against S. lutea before and after treatment with alkaline phosphatase. Fractions numbered 61—250 are combined and concentrated to dryness; yield, 52 g. of material containing clindamycin-3-nucleotides. This material is labeled	10
are tested for activity against S. Intea before and after treatment with alkaline phosphatase. Methanol fraction 2 is concentrated to dryness; yield, 12.63 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.17". Methanol fraction 3 is concentrated to dryness; yield, 0.7 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.2". Preparations ADA—10.1, ADA—11.1, and ADA—12.2, all prepared as described above, are combined as preparation ADA—37.1 (64.7 g.). This preparation containing clindamycin-3-nucleotides is purified further by counter double current distribution A portion (21 g.) of preparation ADA—37.1, described above, is dissolved in 100 ml. of upper and 100 ml. of the lower phase of a solvent system consisting of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase are collected in 50 ml. fractions. A total of 100 transfers are run. The collected fractions and the material in the CDCD tubes are analyzed for S. Intea activity before and after treatment with alkaline phosphatase. Using the same conditions as above, two additional CDCD distributions are run, each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: Pool II Lower-phase collector—Fractions numbered 5—35. Pool I from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.2", and ADA—39.3 and ADA—39.3 comist of essentially pure clindamycin-3-nucleotides by Chromatography The clindamycin-3-nucleotides by use of DEAE—"Sephader" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is aga	15	Fractions W—2 and W—3, described above, and fractions numbered 1—60 from the above Amberlite XAD—2 column, are combined and passed again over the same "Amberlite " (Registered Trade Mark) XAD—2 column, which is first regenerated with 15 liters of water (W—1 obtained as described above) and then clutted with 5 liters of absolute methanol. Three cuts are made, i.e. methanol fraction 1=1	15
Preparations ADA—10.1, ADA—11.1, and ADA—11.2, all prepared as described above, are combined as preparation ADA—37.1 (6.47 g.). This preparation containing clindamycin-3-nucleotides is purified further by counter double current distribution as described below. 2) Counter Double Current Distribution A portion (21 g.) of preparation ADA—37.1, described above, is dissolved in n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase are collected in 50 ml. fractions. A total 100 transfers are run. The collected fractions and the material in the CDCD mibes are analyzed for S. lutea activity before and after treatment with alkaline phosphatase. Using the same conditions as above, two additional CDCD distributions are run, each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: Pool I Lower-phase collector—Fractions aumber 20—50. Pool II Lower and upper-phase remaining in the CDCD machine. Pool II Lower and upper-phase remaining in the CDCD machine. Pool II Tom all three distributions are concentrated to dryness. The resulting residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by fikration and dried; yield, 7.12 g. This preparation is not pursued further. Pools II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides by continued as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephader" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is al	20	hiter; methanol fraction 2=1 liter; and, methanol fraction 3=3 liters. These fractions are tested for activity against S. lutea before and after treatment with alkaline phosphatase. Methanol fraction 2 is concentrated to dryness; yield, 12.63 g. of material containing clindamycin-3-nucleotides. This preparation is labeled "ADA—11.1". Methanol fraction 3 is concentrated to dryness; yield, 0.7 g. of material containing	20
as described below. 2) Counter Double Current Distribution A portion (21 g.) of preparation ADA—37.1, described above, is dissolved in A portion (21 g.) of preparation ADA—37.1, described above, is dissolved in 100 ml. of upper and 100 ml. of the lower phase of a solvent system consisting of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase are collected in 50 ml. fractions. A total of 100 transfers are run. The collected fractions and the material in the CDCD tubes are analyzed for 5. Iutea activity before and after treatment with alkaline phosphatase. Using the same conditions as above, two additional CDCD distributions are run, each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: Pool II Lower-phase collector—Fractions number 20—50. Pool II Lower and upper-phase remaining in the CDCD machine. Pool III Upper-phase collector—Fractions numbered 5—35. Pool I from all three distributions are concentrated to dryness. The resulting residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further. Pool II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea after treatment with alkaline phosphatuse. The presence of clindamycin after phosphatase treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-Nucleotides by Chromatography The clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g	25	Preparations ADA—10.1, ADA—11.1, and ADA—11.2, all prepared as described above, are combined as preparation ADA—37.1 (64.7 g.). This preparation containing	25
100 ml. of upper and 100 ml. of the lower phase of a solvent system constants of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass counter double current distribution apparatus (CDCD) (100 tubes). After 48 transfers, both the upper and lower phase are collected in 50 ml. fractions. A total of 100 transfers are run. The collected fractions and the material in the CDCD tubes are analyzed for S. Iutea activity before and after treatment with alkaline phosphatase. 101		as described below. 2) Counter Double Current Distribution	
35 S. lutea activity before and after treatment with alkaline phosphatase. Using the same conditions as above, two additional CDCD distributions are run, each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: Pool I Lower-phase collector—Fractions number 20—50. Pool II Lower and upper-phase remaining in the CDCD machine. Pool III Upper-phase collector—Fractions number 65—35. Pool I from all three distributions are concentrated to dryness. The resulting residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further. Pools II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.2", and 2.49 g. of material from Pool III labeled "ADA—39.2", and consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase, and activity against S. lutea after treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephader" (A—25) with water for about one hour. The resin is separated by filtration and scirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin prepared as described above, is added into a glass column and allowed	30	100 ml. of upper and 100 ml. of the lower phase of a solvent system consisting of n-butanol-water (1:1 v/v). The solution is added in the center tubes of an all-glass consists double current distribution apparatus (CDCD) (100 mbes). After 48 transfers.	30
each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made: Pool I Lower-phase collector—Fractions number 20—50. Pool II Lower and upper-phase remaining in the CDCD machine. Pool III Upper-phase collector—Fractions numbered 5—35. Pool I from all three distributions are concentrated to dryness. The resulting residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further. Pools II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase. The presence of clindamycin after phosphatase treatment is shown by TLC (Thiri-layer chromatography). D. Separation of Clindamycin-3-nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is spearated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed	35	are run. The collected fractions and the material in the CDCD tibes are analyzed for S. lutea activity before and after treatment with alkaline phosphatase.	35
Pool II Lower and upper-phase remaining in the CDCD machine. Upper-phase collector—Fractions numbered 5—35. Pool I from all three distributions are concentrated to dryness. The resulting residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further. Pools II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase, and activity against S. lutea after treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin prepared as described above, is added into a glass column and allowed		each using 21 g. of preparation ADA—37.1. In each of the above three distributions the following pools of fractions are made:	
residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further. Pools II and III from all three distributions are treated as above for Pool I; yield, 13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase. The presence of clindamycin after phosphatase treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-Nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin prepared as described above, is added into a glass column and allowed	40	Pool II Lower and upper-phase remaining in the CDCD machine. Pool III Upper-phase collector—Fractions numbered 5—35.	40
13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of essentially pure clindamycin-3-nucleotides as evidenced by inactivity against S. lutea before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase. The presence of clindamycin after phosphatase treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-Nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin prepared as described above, is added into a glass column and allowed	45	residue is dissolved in absolute methanol and this solution is mixed with ether. The resulting precipitate is isolated by filtration and dried; yield, 7.12 g. This preparation is not pursued further.	45
before treatment with alkaline phosphatase, and activity against S. lutea after treatment with alkaline phosphatase. The presence of clindamycin after phosphatase treatment is shown by TLC (Thin-layer chromatography). D. Separation of Clindamycin-3-Nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed		13.6 g. of material from Pool II labeled "ADA—39.2", and 0.49 g. of material from Pool III labeled "ADA—39.3". Preparations ADA—39.2 and ADA—39.3 consist of	50
D. Separation of Clindamycia-3-Nucleotides by Chromatography The clindamycia-3-nucleotides, obtained as described above, are separated into the individual clindamycia-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sephadex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed		before treatment with alkaline phosphatase, and activity against <i>S. lutea</i> after treatment with alkaline phosphatase. The presence of clindamycin after phosphatase treatment is shown by TLC (Thin-layer chromatography).	5 0
dex" (A-25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral. The washed resin is then stirred with 0.5 N aqueous acetic acid for 2 hours, and finally washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed	55 .	D. Separation of Clindamycin-3-Nucleotides by Chromatography The clindamycin-3-nucleotides, obtained as described above, are separated into the individual clindamycin-3-nucleotides by use of DEAE—"Sephadex" (Registered Trade Mark) Chromatography. The resin is prepared by slurrying 500 g. of DEAE—"Sepha-	55
washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed	60	dex" (A—25) with water for about one hour. The resin is separated by filtration and stirred with 0.5 N aqueous sodium hydroxide solution for 2 hours. The resin is again isolated by filtration and washed with water until the pH of the wash is almost neutral.	60
		washed to a neutral pH. The resin, prepared as described above, is added into a glass column and allowed	

	then with 4 l. of 0.1% aqueous solution of tris-(hydroxymethyl)-aminomethane (THAM).	
5	The starting material (ADA—39.2, 13.0 g.) is dissolved in 100 ml. of water. The pH is adjusted to 9.0 with concentrated ammonium hydroxide. This solution is then added on the top of the column. The column is eluted sequentially as follows: 1) 15 l. of 0.05 M THAM acetate (prepared by dissolving 6.05 g. of THAM in 800 ml. of water, adjusting the pH to 8.0 with glacial acetic acid and then adjusting the volume to 1 l.).	5
10	 2) 40 L of 0.1 M THAM acetate buffer, pH 8.0. 3) 20 L of 0.2 M THAM acetate buffer, pH 8.0. 4) 20 L of 0.3 M THAM acetate buffer, pH 8.0. Fractions of 20 ml are collected. The following fractions are obtained from each 	10
15	From 0.05 M buffer, fractions 1—722 From 0.1 M buffer, fractions 723—2920 From 0.2 M buffer, fractions 2921—3985 From 0.3 M buffer, fractions 2985—5000	15
20	Selected fractions are analyzed by testing for activity against S. lutea before and after alkaline phosphatase treatment and by U.V. spectra of the effluent of the column both as it is obtained and at acid pH (ca. 2.0). The following pools are made: Pool I Fractions: 850—965	20
25	Volume: ca. 2300 ml. λ max. (a) U.V.: neutral, pH 7.0 270(3.72) acid, pH 2.0 279(5.40) base, pH 11.0 271(3.72)	25
30	Pool II Fractions: 1240—1535 Volume: ca. 5200 ml.	30
35	U.V.: neutral, pH 7.0 261(11.4) acid, pH 2.0 255(11.25) base, pH 11.0 258(11.25) Pool III Fractions: 1550—1680 Volume: 2600 ml.	35
40	U.V.: neutral, pH 7.0 262(3.60) acid, pH 2.0 262(3.64) base, pH 11.0 261(2.82)	40
45	Pool IV Fractions: 1771—2125 Volume: 7000 ml. Δ max. (a) U.V.: neutral, pH 7.0 254(3.74): sh 278 acid, pH 2.0 254(3.66): sh 278	45
50	acid, pH 2.0 254(3.66): sh 278 base, pH 11.0 264(3.20) (a) Isolation of Clindamycin-3-(5'-Cytidylate) present in Pool 1 by Chromatography	50
55	The column is prepared from 150 ml. of "Amberlite"—Registered Trade Mark XAD—2. Pool I, prepared as described above, is passed over the column at a rate of 6 ml./min. The spent liquor is collected in 116 twenty ml. fractions. All fractions show no U.V. maximum and are discarded. The column is then washed with 900 ml. of water (fractions 117—161). The wash is also discarded. The column is then eluted with 80% aqueous methanol. Fractions are analyzed by U.V. Results follow:	55

	Fraction No.	λ max. (a)	
	162	No U.V. maximum	
	163	No U.V. maximum	
	164	No U.V. maximum	_
5	165	No U.V. maximum	5
•	166	No U.V. maximum	
	167	271 (9.9)	
	168	271 (161.8)	
	169	271 (168.0)	
10	170	271 (61.6)	10
••	171	271 (19,4)	
	172	271 (3.6)	
	173	271 (1.0)	
•	174	271 (0.3)	,,
15	175	271 (0.15)	15

Fractions 167—172 are combined. The solution is evaporated to an aqueous concentrate and freeze-dried; yield, 750 mg. of clindamycin-3-(5' cytidylate). Five hundred mg. of this preparation is dissolved in 5 ml. of methanol and the solution is mixed with ether; yield, 400 mg. of clindamycin-3-(5'-cytidylate), having the following structure:

CH₃

 $R_1 = -CH_2CH_3CH_3$

$$z = -\frac{0}{\rho - 0 - CH_2} \frac{1}{\rho}$$
OH OH

Analytical data

Calctl. for C₂:H₂N₃O₁₂PSCl:
C, 44.48; H, 6.17; N, 9.60; O, 26.39; S, 4.39; Cl, 4.87; P, 4.25.
Found: C, 45.62; 45.86; H, 6.99; 7.63; N, 9.80; S, 3.61; Cl, 3.90; 4.04; P, 3.44.

Molecular weight
Calctl. for C₂:H₄₁N₃O₁₂PSCl: 729.5
Found: 742 (vapor pressure osmometry, in methanol).

Potentiometric titration
In water: pKa' 7.7
eq. wt. 587

Specific Rotation: [α]₂²³, +61° (c, 1, water)

Infrared Spectrum: The infrared spectra in both mineral oil mull and KBr pellet are as follows:

In Mineral Oil Mull

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3340	S	1490	S	992	s
3240	S	1455 (oil)	S	967	M
2930 (oil)	S .	1375 (oil)	S	933	M
2860 (oil)	S	1364 (sh)	S	886	S
2730 (sh)	M	1282	S	853	M
1717	M	1215	. s	805 (sh)	M
1650	S	1095	S	787	S
1610 (sh)	s	1070	S	720 (cil)	S
1520	S	1050	S		

In KBr Pellet

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3400	S	1530 (sh)	M	1045	S
3210	S	1520	S	990	M
3100 (sh)	S	1490	S	965	M
2955	S	1455	M	930	M
2925	S	1395	M	882	M
2870	M	1380	M	850	M
2790	M	1286	M	800 (sh)	M
1640	S	1215	, S	785	M
1615	S	1083	S	700	M
1675 (sh)	M	1065	S		

14	1,270,277		
5	Band intensities in the I.R. spectra, disclosed and "W" respectively, and are approximated in vicinity of the bands. An "S" band is of the same in the spectrum; "M" bands are between one-thir strongest band, and "W" bands are less than one-th These estimates are made on the basis of a percent "(sh)" refers to a "shoulder". U.V. Spectrum: In water at the following pH's:	terms of the backgrounds in the order of intensity as the strongest d and two-thirds as intense as the ird as intense as the strongest band.	5
10	pH 2.0 279 13.1	6 9,600	10
10 .	pH 7.0 269 9.3		
	pH 11.0 271 9.1	* 1_1_1_	
15	Reactions with Enzymes Crude Alkaline Phosphatase Treatment with alkaline phosphatase yields chromatography (silica gel, ethyl acetate-acetone-wat Venom Diesterase Treatment with venom diesterase yields clim	indamycin identified by thin-layer er (8:5:1 v/v)).	. 15
	chromatography (as above). In addition to clin	damycin, cytidine-5'-phosphate is	
20	produced.		20
	(b) Isolation of Clindamycin-3-(5'-A	in Pool II	
	by Chromatograph	у	
25	The column is prepared from 400 ml. of "As XAD—2. Pool II is passed over the column at a flois washed with 4 l. of water. Both spent and aqueo and are discarded. The column is eluted with 80% analyzed by U.V. Results follow:	us wash do not show U.V. maxima	25
	Fraction No.	max. (a)	
		50 No maximum	
30		50 (0.18)	30
		50 (0.46)	
		50 (0.47)	
		50 (230.0)	
		60 (632)	35
35		50 (628)	33
•		io (540) io (405)	
		0 (280)	
		0 (230)	
40		0 (135)	40
		0 (80)	
	24 26	io (55)	
		0 (31.5)	
		0 (26.4)	
45		0 (12.8)	45
		0 (9.0) 0 (5.5)	
		0 (5.5) 0 (3.65)	
		0 (3.65) 0 (2.60)	
50		0 (2.0) 0 (2.0)	50
50		0 (1.55)	
		- \/	

Fractions 15—21 are combined. The solution is mixed with 1500 ml. of acetone. The precipitated material is collected and dried; yield, 2.1 g. of clindamycin-3-(5'-adenylate) having the following structure:

 $R_1 = CH_2CH_2CH_3$

Clindamycin-3-(5'-adenylate) has the following chemical and physical properties:

Analytical data
Calcd. for: C₂,H₄,N₇O₁₁PSC1:
C, 44.63; H, 6.05; N, 13.07; S, 4.28; Cl, 4.72; P, 4.11.
Found: C, 44.77; H, 6.66; N, 12.57; S, 4.65; Cl, 4.38; P, 3.52.

Molecular weight
Calcd. for: C₂,H₄,N₇O₁₁PSC1: 753.5
Found: 726 (vapor pressure osmometry, in methanol)

Potentiometric titration
In water: pKa' 7.6
Eq. wt. 620

Specific rotation: [a']_D²³, +62.9° (c, 1.04, water)
Infrared spectrum: The infrared spectra in both mineral oil mull and KBr pellet are as follows:

BNSDOCID: <GB_____1298296A__I_>

In Mineral Oil Mull

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3300	S	1470 (sh)	S	1065	S
3240	S	1450 (oil)	S	1050	S
2940 (oil)	S	1445 (sh)	S	987	S
2920 (oil)	S	1435 (sh)	s	965	S
2845 (oil)	S	1417	s	927	M
1680 (sh)	S	1373 (oil)	s	885	S
1653	S	1363 (sh)	S	853	M
1635	S	1327	S	817	М
1595	s	1298	S	795	S
1570	S	1245 (sh)	S	717 (oil)	S
1510	S	1215	S		

In KBr Pellet

Band Frequency (cm-1)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3340	S	1637	S	1313	S
3270	S	1593	S	1085	S
3220	s	1570	. M	1065	s
2950	M	1510	M	1045	S
2920	S	1470	M	986	M
2865	М	1453	M	927	М
1682	S	1415	M	885	M
1675	S	1375	М	852	м
1660	S	1325	М	815	M
1650	S .	1295	М	795	M
1645	S	1245 (sh)	M	717	м

	U.V. Spectrum: In v	vater at the fo	llowing pH	's:			
			λ max.	a	e		
		pH 2.0	257	16.76	12,628		
	3	PH 7.0	261	16.67	12,560		
5		pH 11.0	261	16.87	12,711	• •	5
	Reactions with Enzy						
	Crude Alkaline	Phos phatase					
	Treatment with	alkaline ph	osphatase y	ields clinda	mycin identified 1	by thin-layer	
	chromatography (sili	ca gel, ethyl a	cetate-aceto	nc-water (8	: 5 : 1 v/v)).	•	
10	Venom Diestera			•	,		10
	Treatment with	venom diestr	erase yields	clindamycin	and adenosine-5	-phosphate.	10
	In Vivo Activity		•	•		1	
	Clindamycin-3-	(5'-adenylate	does not	oosses in viti	ro antibacterial ac	tivity against	
	S. lutea. However, i	ì is active in	vivo (mice,	S.Q., S. aur	eus) with a CDm	of 30 mg/kg.	
2 .	••		•				
Ĭ5 [°]	(c) Isola	tion of Clind	amycin-3-(5	'-Uridylate)	present in Pool I	II	15
			by Chroma	tography	- -		1,
	The column is	prepared fro	m 150 ml.	of "Amberl	lite" (Registered 7	(rade Mark)	
	XAD—2. Pool III	is passed ove	r the colum	n at a rate	of 10 ml./min. T	he column is	
	washed with 1 l. of	water. Both	the spent	and the acu	ucous wash do no	t show U.V.	
20	maximum. The col	umn is then	eluted wit	h 80% aqu	eous methanol. I	ractions are	20
	analyzed by U.V. Re	sults follow:					20
	•	Fraction	No.	λ τα		•	
		2			aximum		
	:	4		261 (0			
25		2	•	261 (0			25
		6		261 (1			_
		7		261 (2			
	•	8		261 (7			
20		. 10		261 (1			
30		10		261 (2			30
		11		261 (0	J.87)		

Fractions 6—9 are combined. The solution is concentrated to dryness. The residue is dissolved in methanol and this solution is mixed with ether to give a precipitate; yield, 740 mg. of clindamycin-3-(5'-uridylate) having the following structure:

 $R_1 = CH_2CH_2CH_3$

$$Z = -P - 0 - C N_{2} O H$$

$$OH OH OH$$

	Clindamycin-3-(5'-uridylate) has the following chemical and physical properties:	
	Analytical data	
	Calcd. for: C ₂₇ H ₄₄ N ₄ O ₁₂ PSCl:	
	C, 44.27; H, 6.33; N, 7.68; O, 28.49; S, 4.39; Cl, 4.86; P, 4.24.	5
5	Found: C, 44.62; H, 6.19; N, 7.79; S, 4.04; Cl, 4.32; P, 4.22.	,
-	Molecular maight	
	Calcil for: CH., N.O., PSCI: 732.5	
	Found: 764 (vapor pressure osmometry in methanol)	,
	Potentiometric titration	10
10	In water: pKa', 7.6	10
	Eq. wt., 576	
	Specific Rotation	
	$[\alpha]_{D}^{25}$, +79.5° (c, 0.99, water)	
	Tulumal Charles	15
15	The infrared spectra in both mineral oil muli and KBr pellet are as follows:	13

In Mineral Oil Mull

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3330	S	1660 (sh)	S	1060	S
3080	S	1545	S	990	S
2950 (oil)	S	1515	S	885	S
2920 (oil)	S	1455 (oil)	S	850	S
2840 (oil)	S	1375 (oil)	s	810	S
1750 (sh)	м	1260	s	763	S
1680	s	1215	S	720 (oil)	s

In KBr Pellet

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3410	S	1685	S	990	M
3260 (sh)	S	1512	M	865	M
3100	M	1458	M	883	M
2960	S	1380	M	850	M
2920	S	1260	s	808	М
2865	M	1215	S	760	М
2800	M	1085	s	705	M
1700 (sh)	S	1060	8		

	λ max.	а	c	
pH 2.0	261	11.18	8,189	
pH 7.0	262	11.44	8,379	
pH 11.0	262	8.90	6.519	
Reaction with Enzymes	202	0.50	0.313	
_ Crude Alkaline Phosphatase				
Treetment with alkaling phase	mhassa si			
Treatment with alkaline phos	phause yn	elos cimoan	lycin identified by thin-layer	•
chromatography (silica gel, ethyl a Venom Diesterase	cetate-acett	me-water (8	:) : 1 v/v)).	
Treatment with venom diest	erase yieids	cundamycu	and uridine-5'-phosphate.	
In Vivo Activity				
(and any cin-5-() - undylate)	does not p	ossess antiba	cterial activity against Sarcina	1
lutea in vitro. However, it is ac	uve in vivo) (S.Q., mic	x, S. aureus) with a CD ₂₀ of	
37 mg/kg.				
/B 7 1 2 4 600 4				
(d) Isolation of Clind	amycin-3-('-guanylane)	present in Pool IV	
	by Chroma	meranhv		
The column is prepared from	m 200 ml.	of Amberlin	XAD-2. Pool IV is passed	
over the column at a rate of 20	ml./min. T	The column	is washed with 2.1 of more	
Doen spent and aqueous wash si	now no II.	V. maximum	The column is alwest with	
80% aqueous methanol. Fractions	are analyze	d by U.V. R	esults follow:	
Fraction	Na	λ <i>ma</i>	• (a)	
2	110.		r (a) Iximum	
_		140 1111	(*shoulder)	
4		No me	(~enouiger)	
6).56); sh+278	
7		254	260); sh 278	
·		254	740); sh 278	
9		257 /	100); sh 278	
10		254 (1	(00); SE 2/8	
ii		254 (1	68); sh 278	
12		254 (0); sh 278	
13		254 (1	(1.5); sh 278	
· 14		224 (3	(2); sh 278	
		23 4 (1	i.16); sh 278	
Fractions 7—10 are combined.	Phin makenie			
PERCEIONS / IU STP COMPANSO 1				

 $R_1 = CH_2CH_2CH_3$

5	Clindamycin-3-(5'-guanylate) has the following chemical and physical properties: Analytical Data Calcd. for: C ₂ :H ₄ :N,O ₃ :PSCl: C, 43.71; H, 5.85; N, 12.74; O, 25.00; S, 4.16; Cl, 4.61; P, 4.03. Found: C, 43.69; H, 6.34; N, 11.62; S, 3.63; Cl, 4.15; P, 3.81.	5
10	Molecular Weight Calcd. for: C ₂ .H ₄₅ N ₇ O ₁₂ PSCI: 769.5 Found: 750 (vapor pressure osmometry in methanol) Potentiometric Titration In water: pKa', 7.6	10
15	Eq. wt., 721 Specific Rotation [a] ₂ ² , +69° (c, 1.0, water) Infrared Spectrum The infrared spectra in both mineral oil mull and KBr pellet are as follows:	15

In Mineral Oil Mull

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3330	S	1530	S	1050 (sh)	S
3220	S	1457	S	987	M
2920 (oil)	S	1409	M	963	M
2845 (oil)	s .	1375 (oil)	S	925	М
1684	S	1365	S	885	S
1675	S	1315	M	855	M
1635	. S	1250 (sh)	S	795	М
1630	S	1215	S	780	M
1595	S	1080 (sh)	S	717 (oil)	М
1565	S	1065	S	680	M

In KBr Pellet

Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity	Band Frequency (cm ⁻¹)	Intensity
3420	S ·	1570	S	1065	S
3240 (sh)	S	1530	M	1045	S
2950	S	1450	M	985	M
2920	S	1405	M	925	M
2865	S	1380	M	885	M
1685	S	1355	M	855	M
1635	S	1255 (sh)	M ·	800	M .
1630	S	1210	. y S	780	M
1595	š · ··	1080	s	• • •	

U.V. Spectrum: In water at the following pH's:

λ max.
256
277 sh
254
273 sh
259
266 pH 1.0 pH 7.0 pH 11.0

10

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Reaction with Ensymes

Crude Alkaline phosphatase

Treatment with alkaline phosphatase yields clindamycin identified by thin-layer chromatography (silica gel, ethyl acetate-acetone-water (8:5:1 v/v)).

	Venom Diesterase	
	Treatment with venom diesterase yields clindamycin and guanosine-5'-phosphate.	
5	In Vivo Activity Clindamycin-3-(5'-guanylate) does not possess antibacterial activity against Sarcina lutea in vitro. However, it is active in vivo (S.Q., mice, S. aureus) with a CD _m of	5
	26 mg./kg.	
	Example 2	
	Upon substituting the microorganism Streptomyces venezuelae, URRL 3527, for	
	the microorganism S. coelicolor Müller, NRRL 3532, in Example 1, there are obtained the clindamycin-3-nucleotides disclosed in Example 1.	10
10	the children yell-y-indicordes textosce in Example 1.	
	Example 3	
	Upon substituting lincomycin for clindamycin in the fermentation medium of	
	Example 1, there are obtained lincomycin-3-nucleotides wherein the nucleotide moieties	
	are the same as disclosed in Example 1.	
15	Example 4	15
L	Upon substituting 1'-demethyl-clindamycin for clindamycin in the fermentation	
	medium of Example 1, there are obtained 1'-demethyl-clindamycin-3-nucleotides	
	wherein the nucleotide moieties are the same as disclosed in Example 1.	
	E	
20	Example 5 Upon substituting 1'-demethyl-4'-depropyl-4'-pentyl-clindamycin for clindamycin	20
20	in the fermentation medium of Example 1, there are obtained 1'-demethyl-4'-depropyl-	20
	4'-pentyl-clindamycin-3-nucleotides wherein the nucleotide moieties are the same as	
	disclosed in Example 1.	
	Example 6	
25	Upon substituting 4'-depropyl-4'-ethyl lincomycin for clindamycin in the fermentation medium of Example 1, there are obtained 4'-depropyl-4'-ethyl lincomycin-3-	25
	nucleotides wherein the nucleotide moieties are the same as disclosed in Example 1.	
	manies and mine and an american transfer are fair partie as described in Example 1.	
	Example 7	
	Upon substituting 1'-demethyl-1'-ethyl lincomycin for clindamycin in the fermen-	
30	tation medium of Example 1, there are obtained 1'-demethyl-1'-ethyl lincomycin-3- nucleotides wherein the nucleotide moieties are the same as disclosed in Example 1.	30
	montes and the management montes and the same as approved in parampte 1.	
	Example 8	
	Upon substituting 1'-demethyl lincomycin for clindamycin in the fermentation	
35	medium in Example 1, there are obtained 1'-demethyl lincomycin-3-nucleotides wherein the nucleotide moieties are the same as disclosed in Example 1.	
33	the municoduc monocles are the same as disclosed in Example 1.	35
	Example 9	
	Upon substituting celesticetin for clindamycin in the fermentation medium in	
	Example 1, there are obtained celesticetin-3-nucleotides wherein the nucleotide moieties are the same as disclosed in Example 1.	
40	In the following examples, as above, the nucleotide moieties of the compounds	
30	of the examples are the same as disclosed in Example 1, i.e., cytidylate, adenylate,	40
	uridylate and guanylate.	
	Example 10	
45	Lincomycin-3-Nucleotide-Ammonium Sak A lincomycin-3-nucleotide in the zwitterionic form is dissolved in a minimum	
4.7	amount of water and diluted with an equal amount of ethanol. The solution is cooled	45
	in an ice-water bath and then saturated with ammonia was. The solution is raken to	
	dryness at 30°C, under high vacuum. The residue is dissolved in a minimum amount	
	of methanol and diluted with 5 volumes of other to precipitate lincomycin-3-nucleotide	
50	as the ammonium salt.	50
	Example 11 Aqueous Oral Drops	
	A lincomycin-3-nucleotide 100 gm.	
	Propyl paraben 0.25 gm.	
55	Methyl paraben 0.75 gm.	<i>5</i> 5
	Sorbic acid 1.0 gm.	
	Sodium hydroxide, 4 N aqueous q.s. to pH 7.5 Water, deionized q.s. 1000 ml.	
	Water, deionized q.s. 1000 ml	

Example 12 Syrup containing 400

	An aqueous oral preparation containing 400 mg. of a lincomycin-3-nucleotide in each five milliliters is prepared from the following ingredients:	
5	A lincomycin-3-nucleotide 800 gm. Methyl paraben, U.S.P. 7.5 gm. Propyl paraben, U.S.P. 2.5 gm. Sorbic acid 10 gm. Saccharin sodium 6.5 gm.	. 5
10	Glycerin 3000 ml. Tragacanth powder 100 gm. Orange oil flavor 10 gm. F.D. and C. orange dye 7.5 gm. Sodium hydroxide, 4 N agueous g.s. pH 7.5	10
15	Desonized water q.s. 10,000 ml.	15
20	In place of a lincomycin-3-nucleoside in Examples 11 and 12, there can be substituted a 7(S)-chloro-7-deoxylincomycin-3-nucleotide, as well as the water soluble salts of a 7(S)-chloro-7-deoxylincomycin-3-nucleotide, for example, the alkali metal salts including the ammonium salt.	
20	The aqueous formulations of Examples 11 and 12 are particularly useful as pediatric preparations and can be administered orally in the same dosages as lincomycin.	20
	Example 13 Capsules	
25	One thousand two-piece hard gelatin capsules for oral use, each containing 350 mg. of a 1'-demethylclindamycin-3-nucleotide are prepared from the following types and amounts of materials:	25
	A 1'-demethylclindamycin-3-nucleotide 350 gm.	
30	Com starch 50 gm.	•
	Magnesium stearate 25 gm. 25 gm. 25 gm.	30
35	The materials are thoroughly mixed and then encapsulated in the usual manner. The foregoing capsules are useful for the systemic treatment of infection in adult humans by the oral administration of 1 capsule every 4 hours. Using the procedure above, capsules are similarly prepared with a 1'-demethyl-clindamycin-3-nucleotide in 50, 125, 250 and 500 mg. amounts by substituting 50, 125, 250 and 500 Gm. of a 1'-demethyl-clindamycin-3-nucleotide for the 350 Gm. used above.	35
40	Example 14 Tablets	
	One thousand tablets for oral use, each containing 500 mg. of a 1'-demethyl-4'-depropyl-4'-pentyl-clindamycin-3-nucleoside are prepared from the following types and amounts of materials:	40
45	A 1'-demethyl-4'-depropyl-4'-pentyl clindamycin-3-nucleotide 500 gm.	
	Lactose 50 gm. Com starch 65 gm. Magnesium stearate 3 gm. Light liquid petrolatum 3 gm.	45
50	The ingredients are thoroughly mixed and slugged. The slugs are broken down by forcing through a number sixteen screen. The resulting granules are then compressed into tablets, each tablet containing 500 mg. of active material. The foregoing tablets are useful for systemic treatment of infections in adult humans by oral administration of 1 tablet every 4 hours.	50
55	Using the above procedure except for make in a	
	to 200 gm., tablets containing 200 mg. of active material are prepared.	- 55

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Example 15

Parenteral Preparation

A sterile aqueous preparation for intramuscular use, containing in 1 ml. 300 mg. of a celesticetin-3-nucleotide is prepared from the following types and amounts of materials:

A celesticatin-3-nucleotide 300 gm.

Benzyl alcohol 9 gm.
Water for injection, q.s. 1000 ml.

The sterile drug is dispensed in the sterile benzyl alcohol-water vehicle and filled into vials and the vials sealed.

Example 16 Animal Feed

One thousand gm. of a feed mix is prepared from the following types and amounts

15 A 4'-depropyl-4'-ethyl lincomycin
3-nucleotide 20 gm.
Soybean meal 390 gm.
Fish meal 400 gm.
Wheat germ oil 50 gm.
20 Sorghum molasses 140 gm.

The ingredients are mixed together and pressed into pellets.

The composition can be fed to laboratory animals, i.e., rats, mice, guinea pigs, and rabbits for prophylaxis during shipping.

For larger animals the composition can be added to the animal's regular feed in

For larger animals the composition can be added to the animal's regular feed in an amount celculated to give the desired dose of active material.

Example 17

Parenteral Preparation
A sterile aqueous preparation for intramuscular use, containing in 1 ml. 300 mg. of a lincomycin-3-nucleotide is prepared from the following types and amounts of materials:

A lincomycin-3-nucleotide 300 gm.

Benzyl alcohol 9 gm.

Water for injection, q.s. 1000 ml.

The sterile drug is dispensed in the sterile benzyl alcohol-water vehicle and filled into vials and the vials scaled.

Example 18

7-Deoxy-7(S)-Methoxylincomycin Hydrochloride
Part 18—A: Methyl N-acetyl-7-deoxy-7(S)-methoxy-α-thio-lincosaminide

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Pr=n-propyl

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A suspension of 2.35 gms. of methyl 6,7-aziridino-6-deamino-7-deoxy-α-thiolincosaminide (VI) was maintained with stirring in 25 ccs. of methanol. To the suspension was then added 2.04 gms. of acetic anhydride. After stirring at room temperature for one hour the solvent was removed on a rotary evaporator at 40°C./7 mm. The resulting solids were then chromatographed on a 4.8 × 94 cm. column of silica gel using 1MeOH: 10 CHCl, as the solvent system. The weight of the silica was 750 gms. After a forerun of 1000 ml., 50 ml. fractions were collected. Fractions 31—85 were combined, and evaporated to dryness yielding 3.2 gms. of methyl N-acetyl-7(S)-methoxy-7-deoxy-α-thiolincosaminide (VII) as a colorless amorphous solid, having the molecular weight by mass spectrometry of 309, compared with the calculated molecular weight of 309,38.

The starting aziridino compound of formula VI can be obtained by dehydrohalogenation of methyl 7(S)-chloro-7-deoxy-a-thiolincosaminide (Belgian Parent 705,427). The dehydrohalogenation is effected with anhydrous sodium carbonate by heating at reflux in dimethylformamide (Belgian Patent 732,352, British Patent Specification No. 1,258,944).

Part 18—B: Methyl 7-deoxy-7(S)-methoxy-α-thiolincosaminide (VIII) (Methyl 6,8-dideoxy-7-O-methyl-6-amino-1-thio-L-threo-α-D-galacto-octopyranoside)

A solution of 3.2 gms. of methyl 7-deoxy-7(S)-methoxy-\(\alpha\)-thiolincosaminide (VII) in 25 gms. of hydrazine hydrate was heated under gentle reflux with stirring in an oilbath at 145°C. overnight. The solvent was removed from the colorless solution as completely as possible by distillation from an oil-bath at 100°C./15 mm. and finally at high vacuum to give methyl 7-deoxy-7(S)-methoxy-\(\alpha\)-thiolincosaminide as a colorless syrup. The syrup was chromatographed on 750 gms. of silica gel in a 4.8 × 97 cm. column using 1 MeOH: 10 CHCl₂ as the solvent system. After 1.4 fitur forerun, 50 ml. fractions were collected. Fractions 281—600 were pooled and evaporated to dryness yielding 2.06 gms. methyl 7-deoxy-7(S)-methoxy-\(\alpha\)-thiolincosaminide (VIII) which on crystallization from acetonitrile yielded colorless needles having the following characteristics:

m.p. 154—155°C.

[a]_D +260° (c, 0.5634, H₂O)

Anal. Calcd. for C₁₁H₂₁O₂NS:

C, 44.92; H, 7.92; N, 5.24; S, 12.00; OMe, 11.61

Found: C, 45.20; H, 7.96; N, 5.08; S, 12.19; OMe, 11.86

Mol. Wt. calcd.: 267.35

Found (mass spec.): 267

Part 18--C: 7-Deoxy-7(S)-methoxylincomycin hydrochloride

BNSDCCID: <GB_____1298295A__f_>

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To a suspension of 2.7 gms. of trans-1-methyl-4-propyl-L-2-pyrrolidinecarboxylic acid hydrochloride in 90 ccs. acetonitrile was added with stirring 2.89 gms. of triethylamine. The stirring was continued until all of the solid had dissolved; the reaction annue. The surring was continued until all of the solid had dissolved; the reaction mixture was then cooled in an ice-methanol bath to -5° C, when a precipitate of trichlylamine hydrochloride appeared. There was then added 1.78 gms. of isobaryl chloroformate dropwise keeping the temperature of the reaction at -5° to -3° C. Additional triethylamine hydrochloride precipitated and stirring was continued at -5° C for 20 minutes. To the resulting reaction mixture was added 1.74 gms. of methal -5° C for -5° C methal -5° 5 methyl 7-deoxy-7(S)-methoxy-r-thiolincosaminide (VIII), dissolved in 10 ccs. of water. As the solids dissolved, the temperature rose to about 0°C. and stirring was continued 10 for 2 hours, without further icing the cooling bath. The solvent was then removed on a rotary evaporator at 40°C./15 mm. to a brown viscous residue. This was dissolved in dilute hydrochloric acid and the solution (pH 2) extracted twice with chloroform and the combined extracts washed once with water. The aqueous phase containing the wash water was adjusted to pH 11 with sodium hydroxide (50% aqueous solution), saturated with sodium chloride and extracted 3 times with chloroform. The combined chloroform 15 extracts were dried over anhydrous sodium sulfate and taken to dryness yielding 1.76 gms, of a tan amorphous solid. The tan amorphous solid was chromatographed on 750 gms, of silica gel in a 4.8 × 94 cm. column using 1 MeOH:15 CHCl, as the solvent system. After 1.3 liters of forerun, 50 ml. fractions were collected. Fractions 60 20 to 80 were pooled and taken to dryness yielding 7-deoxy-7(S)-methoxylincomycin free base as an almost coloriess syrup. This free base was taken up in diffute aqueous HCl and the resulting solution filtered and freeze-dried yielding 801.4 mg. of 7-deoxy-7(S)methoxylincomycin hydrochloride as a colorless amorphous solid having the following characteristics: 25

[a]₉ +117° (c, 0.9626, H₂O)

Anal. Calcd. for C₁, H₂O₄N₂S.HCl:
C, 49.93; H, 8.16; N, 6.13; S, 7.02

Found (corrected for 4.14%, H₂O)
C, 49.44; H, 7.99; N, 6.20; S, 6.48

Mol. Wt. Calcd. for anhydrous free base: 420.57

Found (Mass spec.): 420

The 7-deoxy-7(S)-methoxylincomycin thus produced can be subjected to the processes of the present invention to yield the corresponding novel 3-nucleotides of this invention.

Starting materials for the present invention may be prepared as follows:-

Preparation 1A

Alternative Method for Producing Methyl 7-Deoxy-7(S)-Methoxy-α-Thiolincosaminide
(VIII) Methyl N-acetyl-6,7-aziridino-6-deamino-7-deoxy-2,3,4-tri-0-acetyl-α-thiolincosaminide (X)

To a solution of 2.0 gms. of methyl 6,7-aziridino-6-deamino-7-deoxy- α thiolincosaminide (VI) in 20 ccs. of pyridine was added with stirring 10 ccs. of acetic anhydride and the reaction mixture left overnight at room temperature. The volatile material was removed as completely as possible from the reaction mixture on a rotary evaporator at 40°C./7 mm., finally at high vacuum, to a colorless solid. The resulting solid was dissolved in chloroform, stirred with aqueous cadmium chloride to remove the pyridine, filtered and the chloroform layer washed twice with water, and dried over anhydrous sodium sulfate. On removal of the solvent on the rotary evaporator at 40°C./7 mm.

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methyl N - acetyl - 6,7 - aziridino - 6 - deamino - 7 - deoxy - 2,3,4 - tri - 0 - acetyla-thiolincosaminide (X) was obtained as a colorless crystalline solid, weight 3.1 gms. Recrystallization from ethyl acetate-Skellysolve B (technical hexane) gave colorless prismatic needles having the following characteristics:

5 m.p. 173.5—175°C.
[α]_D +222° (c, 0.912, CHCl₃)

Anal. Calcd. for C₁, H₂₅O.NS:

C, 50.61; H, 6.25; N, 3.47; S, 7.95

Found: C, 50.43; H, 6.33; N, 3.41; S, 8.31

Mol. Wt. calcd.: 403.45

From (Manuel 2)

Found (Mass spec.): 403

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Preparation 1
Methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-methoxy-cthiolinessaminide (XI)

thiolincosaminide (XI)

A mixture of 5 gms. of methyl N-acetyl-2,3,4-O triacetyl-6,7-aziridino-6-deamino-7-deoxy-a thiolincosaminide (X), 50 ccs. methanol, and 5 ccs. glacial acetic acid was heared under gentle reflux in an oil bath at 130°C. for six hours. The solvent was removed from the colorless solution at 40°C/7 mm. on a rotary evaporator yielding a pale yellow syrup which crystallized. The crystals were taken up in methylene chloride solution, washed with saturated aqueous sodium bicarbonate, then with water and then dried over anhydrous sodium sulfate. Removal of the solvent as above gave methyl N-acetyl-2,3,4-tri-0-acetyl-7(S)-methoxy-7-deoxy-a-thiolincosaminide XI) as colorless crystals (5.31 gms.). Crystallization from ethyl acetate-Skellysolve B gave fine colorless needles having the following properties:

m.p. 235—236°C.
[a]_p +205° (c, 0.9952, CHCl₃)
Anal. Calcd. for C_{1p}H₈,O_pNS:

C, 49.64; H, 6.71; N, 3.22; S, 7.36; OMe, 7.13

Found: C, 49.77; H, 6.92; N, 3.65; S, 7.90; OMe, 7.38

Mol. Wt. calcd.: 435.49

Found (Mass spec.): 435

On hydrazinolysis by the procedure of Part 18—B there is obtained methyl 7-deoxy-7(S)-methoxy-a-thiolincosaminide (VIII).

Preparation 2A

Methyl N acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-methoxy-α-thiolincosaminide (XI) and

Methyl N-acetyl-2,3-di-O-acetyl-7-deoxy-7(S)-methoxy-α-thiolincosaminide (XII)

To 26.61 gms. of methyl N-acetyl-7-deoxy-7(S)-methoxy-α-thiolincosaminide (VII) in 100 ccs. of pyridine there was added 50 ccs. of acetic anhydride with stirring and the reaction mixture allowed to stand overnight at room temperature. The volatile materials were then removed by distillation on a rotary evaporator at 40°C./7 mm. and finally under high yacuum. The residue was dissolved in chloroform and washed with saturated aqueous sodium bicarbonate. The aqueous layer was washed with chloroform and the combined chloroform extracts stirred with aqueous cadmium chloride to remove the pyridine.

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5	The precipitate was filtered off and washed well with chloroform and the chloroform layer separated, washed twice with water and dried over anhydrous sodium sulfate. On removal of the solvent on a rotary evaporator at 40°C./7 mm. a pale yellow syrup which crystallized on standing was obtained. On recrystallization from ethyl acetate-Skellysolve B, the product was obtained as small colorless, flattened needles, and had the following characteristics:	5
10	m.p. 245 — 247 °C. [α] _b +202° (c, 0.7142, CHCl ₃) Anal. Calcd. for C ₁ ,H ₂ ,O ₄ ,NS: C, 49.64; H, 6.71; N, 3.22; S, 7.36; OMe, 7.13	10
	Found: C, 49.24; H, 6.75; N, 3.34; S, 7.52; OMe, 7.17 Mol. WL calcd.: 435.49 Found (Mass spec.): 435	
15	The above material by Craig countercurrent distribution using as a solvent system 1 EtOH: 1H ₂ O: 1 EtOAc: 1 cyclohexane was shown to contain 70% of methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-methoxy-α-thiolincosaminide (XI) and 30% of methyl N-acetyl-2,3-di-O-acetyl-7-deoxy-7(S)-methoxy-α-thiolincosaminide (XII). After 500 transfers, fractions from tubes 225—310 were pooled (K value 1.14) and Stellyndyn R gayr	15
20	evaporated to dryness and on recrystalization from ethyl acetae-sensions of gardenethyl N-acetyl-2,3,4-tri-0-acetyl-7-deoxy-7(S)-methoxy-a-thiolincosaminide (XI) as fine colorless needles, identical with the product of Preparation 1—B. Exercises from tubes 115—220 (K value 0.59) were pooled and evaporated to	20
25	dryness and on recrystallization from ethyl acetate-Skellysolve B gave methyl N-acetyl-2,3-di-O-acetyl-7-deoxy-7(S)-methoxy-u-thiolincosaminide (XII) as colorless chunky needles having the following characteristics:	25
30	m.p. 189—190°C. [a] _D +275° (c, 1.0188), CHCl ₃) Anal. Calcd. for C _{1c} H ₂ :O ₄ NS: C, 48.84; H, 6.92; N, 3.56; S, 8.15; OMe, 7.89 Found: C, 48.71; H, 7.11; N, 3.93; S, 7.96; OMe, 7.98 Mol. Wt. calcd.: 393.46 Found (Mass spec.): 393	30
35	Preparation 2B Acetylation of methyl N-acetyl 2,3-di-O-acetyl-7-deoxy-7(S)- methoxy-a-thiolincosaminide (XII) To a solution of 200 mg. of methyl N-acetyl-2,3-di-O-acetyl-7-deoxy-7(S)- methoxy-a-thiolincosaminide (XII) in 20 ccs. of pyridine was added 10 ccs. of acetic	35
40	anhydride with stirring and the reaction mixture left at room temperature overlight. The solvent was removed from the colorless reaction solution on a rotating evaporator at 40°C.7 mm. finally at 40°C./high vacuum. The syrupy residue was dissolved in chloroform, washed with dilute aqueous HCl (1/2 normal), twice with water, with	40
45	sodium sulfate. The solvent was then removed on a rotating evaporator at 40°C/7 mm. yielding methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-methoxy-tr-thiolincosaminide (XI) as a colorless syrup which crystallized on standing. On hydrazinolysis of the products of Preparation 2A and 2B, there is obtained methyl 7-deoxy-7(S)-methoxy-thiolincosaminide (VIII).	45

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Preparation 3A Methyl N-acetyl 6,7-aziridino-6-deamino-7-deoxya-thiolincosaminide (XIII)

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To a suspension of 2.3 gms, of methyl 6,7-aziridino-6-deamino-7-deoxy-a-thio-lincosaminide (VI) in 25 ccs. isopropyl alcohol, there was added with stirring 2.04 gms. acetic anhydride. Most of the solid appeared to go into solution to be replaced by new solid. The reaction mixture was stirred or any other receipts at room temperature, then filtered and the receipts are reached with interest and the receipts at room temperature. the residue washed with isopropyl alcohol and dried in a vacuum oven at 60°C/15 mm. There was obtained 2.28 gms. of methyl N-acetyl-6,7-aziridino-6-deamino-7-deoxy-athiolincosaminide as colorless platelets having the following properties:

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m.p. 145°C.
[a]_n +253° (c, 0.7916, H₂O)

Anal. Calcd. for C₁₁H₁₀O₂NS:

C, 47.63; H, 6.91; N, 5.05; S, 11.56

Found: C, 47.57; H, 6.71; N, 5.23; S, 11.29

Mol. Wt. calcd.: 277.34

Found (Mass spec.): 277

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Preparation 3—B

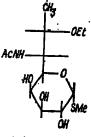
Methyl N-acetyl-7-deoxy-7(S)-methoxy-a-thiolincosaminide (VII)

On treating methyl N-acetyl 6,7-aziridino-6-deamino-7-deoxy-a-thiolincosaminide (XIII) with methanol and acetic acid under reflux, there is obtained methyl N-acetyl-7-deoxy-7(S)-methoxy-a-thiolincosaminide (VII) identical with the product of Part

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Preparation 4—A
7-Deoxy-7(S)-Ethoxylincomycin Hydrochloride
Methyl N-acetyl-7-deoxy-7(S)-ethoxy-α-thiolincosaminide (XIV)



XIY

On treating the methyl N-acetyl-6,7-aziridino-6-deamino-7-deoxy-a-thiolincosaminide (XIII) with ethanol and acetic acid under gentle reflux, there is obtained methyl-N-acetyl-7-deoxy-7(S)-ethoxy-1-thio-a-lincosaminide (XIV) as a syrup having the molecular weight by mass spec. of 323 compared with the calculated molecular weight of 323.41.

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Preparation 4-B

Methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-ethoxy-a-thiolincosaminide (XVI) and Methyl N-acetyl-7-deoxy-7(S)-ethoxy-2,3-di-O-acetyl-a-thiolincosaminide (XVI)

On treating the methyl N-acetyl-7-deoxy-7(S)-ethoxy-a-thiolincosaminide (XIV) with acetic anhydride and pyridine by the process of Preparation 2A there is obtained methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-O-ethyl-a-thiolincosaminide (XV) together with a minor amount of N-acetyl-2,3-di-O-acetyl-7-deoxy-7(S)-ethoxy-a-thiolincosaminide (XVI). The products (isolated on a Craig in 500 transfers using ethanol: water: ethyl acetate: cyclohexane (1:1:1:1) as the solvent system) are characterized as follows: as follows:

Mixture: m.p. 197—199°C.

[α]_b +247° (c, 0.665, CHCl₃)

And. Calcd. for C_{1n}H₃₁O_nNS:

C, 50.76; H, 6.95; N, 3.12; S, 7.13; OEt, 10.02

Found: C, 50.42; H, 7.07; N, 3.18; S, 7.37; OEt, 11.85

Pure XV (K=1.59): m.p. 254—255°C.

[α]_b +199° (c, 0.8638, CHCl₃

Anal. Calcd. for C_{1n}H₃₁O_sNS:

C, 50.76; H, 6.95; N, 3.12; S, 7.13; OEt, 10.02

Found: C, 50.75; H, 7.06; N, 3.37; S, 7.31; OEt, 10.25

Mol. Wt. calcd.: 449.52

Found (Mass spec.): 449

Pure XVI (K=0.87): m.p. 215.5—216.5°C.

[α]_b+261° (c, 1.0448, CHCl₃)

Anal. Calcd. for C₁₁H₃₂O_sNS

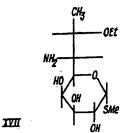
C, 50.11; H, 7.17; N, 3.44; S, 7.78

Found: C, 50.17; H, 7.30; N, 3.50; S, 7.62

Mol. Wt. calcd.: 407.48

Mol. Wt. calcd.: 407.48 Found (Mass spec.): 407

Preparation 4—C
Methyl 7-deoxy-7(S)-ethoxy-a-thiolincosaminide (XVII)



On subjecting the products of Preparation 4B, that is the mixture, the pure XV or the pure XVI to hydrazinolysis, there is obtained methyl 7-deoxy-7(S)-ethoxy-a-thiolincosaminide (XVII) having the following characteristics:

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m.p. 194—196°C.
[a]_D +252° (c, 0.7438, H₂O)

Anal. Calcd. for C₁₁H₂₂O₃NH:
C, 46.95; H, 8.24; N, 4.98; S, 11.40

Found: C, 46.66; H, 8.09; N, 5.26; S, 11.33

Mol. Wt. calcd.: 281.37

Found (Mass spec.): 281

Preparation 4—D 7-Deoxy-7(S)-ethoxylincomycin hydroheloride (XVIII)

Me=methyl Et =ethyl Pr =propyl

Following the procedure of Part 18—C, methyl 7-deoxy-7(S)-ethoxy-α-thiolincosaminide (XVII) is converted to 7-deoxy-7(S)-ethoxylincomycin hydrochloride having the following characteristics:

m.p. colorless amorphous solid
[α]n + 109° (c, 0.9824, H₂O)

Anal. Calcd. for C₂₀H₁₀O₄N₂S.HCl:

C, 50.99; H, 8.35; N, 5.95; Cl, 7.53; S, 6.81; OEt, 9.57

Found (corrected for 5.07% water)

C, 50.54; H, 8.19; N, 5.63; Cl, 7.61; S, 6.95; OEt, 10.16

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Preparation 5A

Methyl N-acetyl-7-deoxy-7(S)-propoxy-a-thiolincosaminide (XIX), methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-propoxy-a-thiolincosaminide (XX), and methyl N-acetyl-7-deoxy-7(S)-propoxy-2,3-di-O-acetyl-a-thiolincosaminide (XXI)

0Pr ACNH XXI ÒAC

On treating the methyl N-acetyl-6,7-aziridino-6-deamino-7-deoxy-\alpha-thiolincosaminide (XIII) with propanol and acetic acid under gentle reflux, there is obtained methyl N-acetyl-7-deoxy-7(S)-propoxy-\alpha-thiolincosaminide (XIX) from which on acetylation with acetic anhydride in pyridine by the procedure of Part 22—B, there is obtained methyl N-acetyl-2,3,4-tri-O-acetyl-7(S)-propoxy-7-deoxy-\alpha-thiolincosaminide (XX) containing a minor amount of methyl N-acetyl-2,3-di-O-acetyl-7(S)-propoxy-7-deoxy-\alpha-thiolincosaminide (XXI) having the following characteristics:

Mixture: m.p. 240—242°C. [α]_p +207° (c, 0.9054, CHCl_a) 4nal. Calcd. for C_aH₃₃O_aNS: C, 51.81; H, 7.17; N, 3.03; S, 6.92 Found: C, 51.41; H, 7.33; N, 3.16; S, 6.92 15

Found: C, 51.41; H, 7.35; N, 3.16; S, 6.92
Mol. Wt. calcd.: 463.60
Found (Mass spec.): 463
Pure XX: m.p. 241.5—242.5°C.

[a]_p +193° (c, 0.9254, CHCl₃)
Anal. Calcd. for C₂H₂₃O₂NS:
C, 51.81; H, 7.17; N, 3.03; S, 6.92
Found: C, 51.77; H, 7.02; N, 3.37; S, 6.84
Mol. Wt. calcd.: 463.60
Found (Mass Spec.): 463

25 Found (Mass Spec): 463

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Preparation 5—B Methyl 7-deoxy-7(S)-propoxy-a-thiolincosaminide (XXII)

On hydrazinolysis of the above products (Preparation 5A) there is obtained methyl 7-deoxy-7(S)-propoxy-α-thiolincosaminide (XXII).

Preparation 5C 7-Deoxy-7(S)-propoxylincomycin hydrochloride (XXIII)

On acylation with trans-1-methyl-4-propyl-L-2-pyrrolidine-carboxylic acid by the procedure of Part 18—C, there is obtained 7-deoxy-7(S)-propoxylincomycin hydrochloride (XXIII).

Preparation 6—A
Methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)isopropoxy-a-thiolincosaminide (XXIV)

Following the procedure of Preparation 1B substituting the methanol by isopropyl alcohol, there is obtained methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-isopropoxy-n-thiolincosaminide (XXIV) having the following characteristics:

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m.p. 253—254°C. $[\alpha]_D + 192^\circ$ (c, 0.535, CHCl₃) Anal. Calcd. for $C_\infty H_{33} O_D NS$: C, 51.81; H, 7.17; N, 3.03; S, 6.92 Found: C, 51.96; H, 7.07; N, 3.19; S, 6.61 Mol. Wt. calcd.: 463.6 Found (Mass spec.): 463

Preparation 6—B
Methyl 7-deoxy-7(S)-isopropoxy-α-thiolincosaminide (XXV)

iPr=Isopropyl
On hydrazinolysis of compound XXIV (Preparation 6A) there is obtained methyl
7-deoxy-7(S)-isopropoxy-a-thiolincosaminide having the following characteristics:

m.p. 213°C.
[\alpha]_n + 225° (c, 0.376, H₂O)

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Anal. Calcd. for C₁₂H₂₅O₂NS:

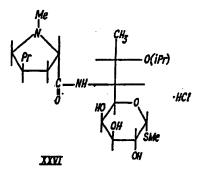
C, 48.79; H, 8.53; N, 4.74; S, 10.86

Found: C, 48.52; H, 8.55; N, 5.26; S, 10.84

Mol. Wt. calcd.: 295.40

Found (Mass spec.): 295

Preparation 6—C
7-Deoxy-7(S)-isopropoxylincomycin hydrochloride (XXVI)



Following the procedure of Part 18—C, compound XXV (Preparation 6B) is converted to 7-deoxy-7(S)-isopropoxylincomycin hydrochloride having the following characteristics:

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35 m.p. amorphous
[\$\alpha\$_p + \$1\circ\$ (c, 0.898, H_0)

Anal. Calcd. for C_1, H_0, O_1, N_3, S. HC1:

C, 51.99; H, 8.52; N, 5.78; S, 6.61; Cl, 7.31

Found (corrected for 4.36%, H_0)

C, 51.72; H, 8.33; N, 5.59; S, 6.35; Cl, 7.29

Mol. Wt. calcd. (free base): 448.62

Found: 448 5 5 Found: 448 Activity: about the same as lincomycin Preparation 7—A
Methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)cyclohexyloxy-a-thiolincosaminide (XXVII) 10 10 OCyclohexyl ACNH XXVII Following the procedure of Preparation 1B substituting the methanol by cyclohexanol, there is obtained methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-cyclohexyl-oxy-a-thiolincosaminide (XXVII) having the following characteristics: 15 15 m.p. 266—268°C.

[a]_D +163° (c, 1.055, CHCl₃)

Anal. Calcd. for C₂,H₃,O_nNS:

C, 54.85; H, 7.41; N, 2.78; S, 6.37

Found: C, 54.93; H, 7.53; N, 2.87; S, 6.65

Mol. Wt. calcd.: 503.61

Found (Mass spec.): 503 20 20 Preparation 7—B
Methyl 7(S)-cyclohexyloxy-7-deoxy-a-thiolincosaminide (XXVIII) 25 25

OCyclohexyl XXVIII

On hydrazinolysis of compound XXVII (Preparation 7A), there is obtained methyl 7(S)-cyclohexyloxy-7-deoxy-n-thiolincosaminide (XXVIII).

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Preparation 7—C 7(S)-cyclohexyloxy-7-deoxylincomycin hydrochloride

Following the procedure of Part 18—C, methyl 7(S)-cyclohexyloxy7-deoxy-a-thiolincosaminide (XXVIII) is converted to 7(S)-cyclohexyloxy-7-deoxylincomycin hydrochloride.

Preparation 8—A

Methyl N-acetyl-7-deoxy-7(S)-2'-hydroxyethoxy-1-thiolincosaminide (XXIX) and methyl N - acetyl - 2,3,4 - tri - O - acetyl - 7(S) - 2' - acetoxyethoxy - 7 - deoxyn-thiolincosamide (XXX)

Following the procedure of Part 18—A substituting the methanol by 2-hydroxy-ethanol, there is obtained methyl N-acetyl-7-deoxy-7(S)-2'-hydroxyethoxy-a-thio-lincosaminide (XXIX) which when acylated by the procedure of Preparation 2A but with heating on a steam bath to produce the fully acylated product gives methyl N-acetyl-2,3,4-tri-0-acetyl-7(S)-2'-acetoxy-ethoxy-7-deoxy-athiolincosaminide having the following characteristics:

m.p. 223—225°C.
[a]_D +172° (c, 1.010, CHCl₃)

Anal. Calcd. for C₂₁H₄₂O₁₁NS:

C, 49.69; H, 6.55; N, 2.76; S, 6.32

Found: C, 49.56; H, 6.63; N, 2.90; S, 6.63

Mol. Wt calcd.: 507.55

Found: 507

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Preparation 8-B Methyl 7-deoxy-7(S)-2'-hydroxyethoxy-a-thiolincosaminide (XXXI)

On hydrazinolysis of methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-2'-acetoxy-ethoxy- α -thiolincosaminide (XXX), there is obtained methyl 7-deoxy-7(S)-2'-hydroxy-ethoxy- α -thiolincosaminide having the following characteristics: 5

m.p. 178.5—179.5°C.

[a]_p +243° (c, 0.662, H₂O)

Anal. Calcal. for C₁₁H₂₁O₄NS:

C, 44.43; H, 7.80; S, 10.78; N, 4.71

Found: C, 44.40; H, 7.99; S, 10.51; N, 4.60

Mol. Wt. calcal.: 297.37 10

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Found (Mass spec.): 297

Preparation 8-C Methyl 7-deoxy-7(S)-2'-hydroxyethoxylincomycin hydrochloride (XXXII)

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Following the procedure of Part 18—C, methyl 7-deoxy-7(S)-2'-hydroxyethoxy- α -thiolincosaminide (XXXI) is converted to 7-deoxy-7(S)-2'-hydroxyethoxy-lincomycin hydrochloride having the following characteristics:

m.p. amorphous
[\$\alpha_{\text{p}}\$ 105° (c, 1.102, H₂O) \\
Anal. Calcd. for C₂₂H₃₄O₇N₂S.HCi:
C, 49.32; H, 8.07; N, 5.75; S, 6.58; Cl, 7.28
Found (corrected for 2.11% H₂O)
C, 49.61; H, 7.85; N, 5.54; S, 6.46; Cl, 7.26
Mol. Wt. calcd. (free base): 450.59
Found (Mass spec.): 450
Activity: about 1/3 lincomycin 20

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Preparation 9—A

Methyl N-acetyl-7-deoxy-7(S)-2'-methoxyethoxy-α-thiolincosaminide (XXXIII) and methyly N - acetyl - 2,3,4 - tri - O - acetyl - 7 - deoxy - 7(S) - 2' - methoxyethoxy-α-thiolincosaminide (XXXIV)

Following the procedure of Part 18.—A but substituting the methanol by 2-methoxyethanol, there is obtained methyl N-acetyl-7-deoxy-7(S)-2'-methoxyethoxy- α -thiolincosaminide (XXXIII) which on acetylation by the procedure of Preparation 2A but with heating on a steam bath to produce the fully acetylated product yields methyl N - acetyl - 2,3,4 - tri - O - acetyl - 7 - deoxy - 7(S) - 2' - methoxyethoxy - α - thiolincosaminide (XXXIV) which is characterized as follows:

m.p. 222—223°C.
[a]₁₁ + 177° (c, 1.079, CHCl₃)

Anal. Calcd. for C₂H₁₅O₁₆NS:

C, 50.09; H, 6.94; N, 2.92; S,6.69; OMe, 6.47

Found: C, 50.13; H, 7.00; N, 2.77; S, 6.33; OMe, 7.28

Mol. Wt. calcd.: 479.54

Found (Mass spec.): 479

Preparation 9—R

Preparation 9—B

Methyl 7-deoxy-7(S)-2'-methoxyethoxy-α-thiolincosaminide (XXXV)

On hydrazinolysis of methyl N-acetyl-2,3,4-tri-O-acetyl-7-deoxy-7(S)-2'-methoxy-ethoxy-a-thiolincosaminide (XXXIV), there is obtained methyl 7-deoxy-7(S)-2'-methoxyethoxy-a-thiolincosaminide (XXXV) having the following characteristics:

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Preparation 9—C 7-Deoxy-7(S)-2'-methoxyethoxylincomycin hydrochloride (XXXVI)

Following the procedure of Part 18—C, methyl 7-deoxy-7(S)-2'-methoxyethoxy- α -thiolincosaminide (XXXV) is converted to 7-deoxy-7(S)-2'-methoxyethoxylincomycin hydrochloride having the following characteristics:

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m.p. amorphous
[α]_D +87° (c, 0.575, H₂O)

Anal. Calcd. for C₂₁H₄₀O₁N₂S.HCl:
C, 50.33; H, 8.25; N, 5.59; S, 6.40; Cl, 7.08

Found: (corrected for 4.17% H₂O)
C, 50.47; H, 8.60; N, 5.26; S, 5.86; Cl, 7.50

Mol. Wt. calcd. (free base): 464.62

Found (Mass spec.): 464

Activity: about 1/3 lincomycin 10

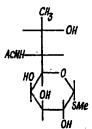
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Preparation 10 Methyl 7-Deoxy-7(S)-Hydroxy-a-Thiolincosaminide (XXXVII) (Methyl 6-Amino-6,8-Dideoxy-L-Threo-a-D-Galacto-Octopyranoside)

 $\begin{tabular}{lll} A: & Methyl & N-acetyl-7-deoxy-7(S)-hydroxy-n-thiolincosaminide & (XXXVIII) & (methyl 6-acetamido-6,8-dideoxy-L-threo-n-D-galacto-octopyranoside) & (Methyl N-acetyl-7-deoxy-7(S)-hydroxy-n-thiolincosaminide & (XXXVIII) & (methyl 6-acetamido-6,8-dideoxy-L-threo-n-D-galacto-octopyranoside) & (Methyl N-acetyl-7-deoxy-7(S)-hydroxy-n-thiolincosaminide & (Methyl N-acetyl-7-deoxy-$ 20



XXXVIII

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To a solution of 2.45 gms. of methyl 6,7-aziridino-6-deamino-7-deoxy-a-thiolincosaminide (VI) in 25 ccs. of water was added 2.04 gms. of acetic anhydride and the solution left at room temperature overnight. The solution was then taken to dryness on a rotary evaporator at 40°C./7 mm. to give a colorless syrup which was chromatographed on 750 gms. of silica gel in a 4.8 × 98 cm. column using 1MeOH:7 CHCl, as the solvent system. After a forerun of 550 ml., 50 ml. fractions were collected. Fractions 90 to 160 were pooled and taken to dryness to give 2.3 gms. of methyl N-acetyl 7-deoxy-7(S)-hydroxy-a-thiolincosaminide as a colorless solid which crystallized from methanol as coloriess rods having the following characteristics:

m.p. 218-219°C. 10 | [a]_D +260° (c, 1.0296, H₂O) Anal. Calcd. for C₁₁H₂₁O₂NS: C, 44.73; H, 7.17; N, 4.74; S, 10.86 Found: C, 44.89; H, 7.02; N, 5.16; S, 10.64 Mol. Wr. calcd.: 295.36 Found (Mass spec.): 295 15

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B: Deacetylation

The crystallized material from Part 28—A was combined with the mother liquors and taken to dryness on a rotary evaporator at 40°C./7 mm. to give 2.01 gms. solid and taken to dryness on a rotary evaporator at 40 C/7 min. to give a substance with which was heated overnight under gentle reflux with 40 cs. of hydrazine hydrate with stirring. The solvent was removed from the colorless solution on a rotary evaporator at 7 mm. pressure in an oil bath at 120°C. The resulting colorless crystalline residue on recrystallization from methanol gave methyl 7-deoxy-7(S)-hydroxy-a-thiolincosaminide (XXXVII) as colorless platelets having the following characteristics:

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m.p. 211-212°C. 25 m.p. 211—212 [a]_p +280° (c, 0.7728, H.O) Anal. Calcd. for C.H₁,O₂NS: C, 42.67; H, 7.56; N, 5.53; S, 12.66 Found: C, 42.81; H, 7.69; N, 5.85; S, 12.73 Mol. Wr. calcd.: 253.32 30 Found (Mass spec.): 253 Preparation 11

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Methyl N-Acetyl-2,3,4-Tri-O-Acetyl-7(S)-ethoxy-7-Deoxy-a-Thiolincosaminide (XV) and Methyl N - Acetyl - 2,3,4 - Tri - O - Acetyl - 7(S) - Acetoxy - 7 - Deoxy-a-Thiolincosaminide (XXXIX)

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Following the procedure of Preparation 1B but substituting the methanol by Following the procedure of Preparation 1B but substituting the methanol by ethanol, there is obtained methyl N-acetyl-2,3,4-tri-O-acetyl-7(S)-ethoxy-7-deoxy-a-thiolincosaminide (XV) identical with the product of Part 22—B and a minor amount of N-acetyl-2,3,4-tri-O-acetyl-7(S)-acetoxy-7-deoxy-a-thiolincosaminide (XXXIX) which can be separated by Craig countercurrent distribution using 1 EtOH: 1 H₂O: 1 EtOAc: 1.5 cyclohexane as the solvent system in 500 transfers. The minor component (XXXIX) was obtained from tubes numbers 140—200 (K=0.52), and the major component (XXXIX) crystallized from ethylv acetate as colorless needles having the following (XXXIX) crystallized from ethyly acetate as colorless needles having the following characteristics:

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m.p. 312-313°C

m.p. 512—513—6. [\(\alpha\)_p + 182° (c, 0.5898, CHCl₂) Anal. Calcd. for C₁₂H_{2n}O₁₀NS: C, 49.22; H, 6.31; N, 3.02; S, 6.92 Found: C, 49.17; H, 6.51; N, 3.08; S, 6.81

Mol. Wt. calcd.: 463.50 Found (Mass spec.): 463

On subjecting the minor component to hydrazinolysis there is obtained methyl 7-deoxy-7(S)-hydroxy-a-thiolincosaminide (XXXVII) identical with the product of

Example 19

Part 19-A: 2'-Hydroxyethyl N-acetyl-2'-2,3,4-tetra-O-acetyl-7-O-methyl 1-thio-alincosaminide (XIL)

2'-Hydroxyethyl 1-thio-a-celestosaminide (1.0 gm.) (Example 3 of U.S. Patent 3,255,174) was left overnight in solution in pyridine (25 ccs.) and acetic anhydride (12 ccs.). Removal of the solvent in vacuo gave a colorless oil which was dissolved in chloroform, washed with water, dilute aqueous hydrochloric acid, water, saturated aqueous sodium bicarbonate, water and dried over anhydrous sodiuf sulfate. Solvent 15 removal in vacuo gave a syrup (2.03 gms.) which on crystallization from ethyl acetate-Skellysolve B yielded 2'-hydroxyethyl N-acetyl-2'-2,3,4-tetra-O-acetyl-7-O-methyl-1-thio-a-lincosaminide (Formula XL) in squat, colorless prisms, m.p. 143—144°C. 20 Skellysolve B is a brand of technical hexane.

Anal. Calci. for $C_{01}H_{32}O_{11}NS$: C, 49.68; H, 6.54; N, 2.76; S, 6.32% Found: C, 49.66; H, 6.50; N, 2.91; S, 6.34% $[\alpha]_D + 216^\circ$ (C, 0.7746, CHCl₃) 25

-B: Methyl N-acotyl-2,3,4-tri-O-acetyl-7-O-methyl-1-thio-α- and -β-lincosaminides (XLI and XLII)

A solution of 5.05 gms. (1.62 ccs.) of bromine in 100 ccs. of chloroform was added over approximately 30 minutes from a pressure-equalized dropping funnel under anhydrous conditions to a stirred solution of 10 gms. of 2'-hydroxyethyl N-acetyl-2',2,3,4-tetra-O-acetyl-1-thio-a-celestosaminide prepared by the procedure of Part 30—A in 200 ccs. of chloroform. Initially, the bromine color disappeared immediately; later, a deep orange-red color developed. After stirring for an additional 30 minutes at room temperature, solvent was removed on a rotating evaporator at 40°C./7 mm., giving a yellow-orange syrupy residue. This was redissolved in chloroform, the solvent

removed in vacuo, and the process repeated till the distillate became colorless, leaving a yellowish amorphous residue of 1-bromo-7-O-methyl- β -lincosamine tetraacetate of the formula

The residue was dissolved in 200 ccs. of dry dimethylformamide, 4.5 gms. of thiourea was added, and the reaction mixture (a colorless solution) stirred overnight at room temperature. There were thus formed the isothiouronium salts of the formulas

Without isolating these salts and after cooling in an ice-bath, 100 ccs. of water was added slowly, followed by 8.3 gms. of anhydrous potassium carbonate, 10.6 gms. of sodium bisulfite, and 28 gms. (12.3 ccs.) of methyl iodide. The mixture was stirred vigorously magnetically for 3 hours, the cooling bath being removed after 20 minutes.

Volatile materials were removed in vacuo at 40°C., and finally at 80°C./<1 mm.

Volatile materials were removed in occuo at 40°C., and many at 50°C./ 1 min. The yellow residue was dissolved in a mixture of chloroform and water, the aqueous layer extracted with chloroform, and the combined chloroform extracts were washed twice with water and dried over anhydrous sodium sulfate. Removal of the solvent in occuo gave a colorless amorphous residue (6.48 gms.). Thin-layer chromatography (1 acetone: 1 Skellysolve B) showed a major zone of product with a small zone of slightly higher R₄.

This material was chromatographed on silica gel (1.2 kilos, column dimensions 5.8 × 90 cms.) in the system 1 acetone: 1.5 Skellysolve B. After a 500 cc. forerun, 50 cc. fractions were collected automatically, and elution of materials followed by thin-layer chromatography. Fractions numbers 145—173, inclusive, corresponded to the material of higher R_s, numbers 185—310, inclusive, corresponded to the major product, and numbers 174—184, inclusive, were a mixture of the two.

Removal of solvent in vacuo from combined fractions 145—173, inclusive, gave a colorless syrup (570 mgms.), which on crystallization from ethyl acetate-Skellysolve B yielded methyl N-acetyl-2,3,4-tri-O-acetyl-7-O-methyl-1-thio-α-lincosaminide in small colorless prisms, m.p. 212—213°C. undepressed on a mixture with the sample of Example 31 (Part 31—C), of m.p. 211.5—213°C., and also indistinguishable from it by infrared, nuclear magnetic resonance, and mass spectra, and also by optical rotation. Removal of solvent in vacuo from combined fractions 185—310, inclusive, gave a

Removal of solvent in vacual from combined fractions 163—370, including gave a slightly yellow amorphous solid (4.23 gms.) which on crystallization yielded methyl N-acetyl-2-3,4-tri-O-acetyl-7-O-methyl-1-thio-\(\beta\)-lincosaminide in colorless prisms, m.p. 187—188°C.

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Anal. Calcd. for C<sub>12</sub>H<sub>2</sub>,O<sub>2</sub>NS:
C, 49.64; H, 6.71; N, 3.22; S, 7.36; MeO, 7.13
                                                              M.W. 435.49.
                                    Found: C, 49.73; H, 6.95; N, 3.18; S, 7.64; MeO, 7.41 [α]<sub>D</sub> +24° (c, 0.7484, CHCl<sub>2</sub>) Mol. Wt.: (mass spec., M+) 435
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                      The overall yield of introduction of the —SMe group (i.e. \alpha+\beta-anomers) was 49.2% (6.7% \alpha, 42.5% \beta) with the \alpha/\beta ratio 1:6.35.

The \beta-anomer can be recycled to Part 30—B and thus enhance the overall yield
                        of the more desired a-anomer.
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                        Part 30-C:
                       The procedure of Part 30—B was repeated substituting the methylformamide by hexamethylyphosphoric triamide (Me<sub>2</sub>N)<sub>2</sub>P=O) giving an overall yield of 65.5% (22.7% \alpha, 42.8% \beta) and thus an \alpha/\beta ratio of 1:1.9.
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                                               -D-1: Methyl 7-O-methyl-1-thio-u-lincosaminide (XLVI)
                                                                                                                                                                                                                                                                                          15
                                                                                                                           CH30
                                                                                                             III
                                     The methyl 7-O-methyl-1-thio-a-lincosaminide-tetraacetate (XLI) (1.46 gms.) was
                        dissolved in 50 ccs. of hydrazine hydrate and heated under gentle reflux in an oil-bath
                        at 155°C. for 24 hours. Volatile solvent was then removed as completely as possible by
                        distillation at 110°C./15 mm., giving a colorless crystalline residue which was triturated with anhydrous acetonitrile. The solid was removed by fitnessian and dried. On crystallization from a concentrate of 95% ethanol, 430 mgs. of methyl 7-O-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-thio-a-methyl-1-t
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                                                                                                                                                                                                                                                                                         20
                        lincosaminide hemihydrate (Polymorph I) were obtained as colorless flattened needles,
                        m.p. 126-126.5°C.
                                    Anal. Calcd. for C<sub>10</sub>H<sub>-1</sub>O<sub>2</sub>HS.1/2H<sub>2</sub>O:

C, 43.46; H, 8.03; N, 5.07; S, 11.60; OMe, 11.23

M.W. (anhydrous) 267.35.

Found: C, 43.63; H, 8.30; N, 5.18; S, 11.67; OMe, 11.74
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                                       [\alpha]_D + 263^\circ (c, 0.8284, H<sub>2</sub>O)
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                                     Mol. Wt.: (mass spec., M+) 267
                        Part 19-D-2:
                         The procedure of Part 19—D—1 was repeated except that the crystallization was effected slowly in a more dilute solution in 95% ethanol. Methyl 7-O-methyl-1-
                         thio-a-lincosaminide hemihydrate was obtained as colorless platelets, m.p. 162-163°C.
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                                                                                                                                                                                                                                                                                         35
                         Both polymorphic forms showed identical chromatographic behavior (R<sub>1</sub> 0.2 on silica gel TLC in 1 methanol: 15 chloroform by volume). A mixture melting point of
                        forms I and II gave the following:
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                                                                        I and I
                                                                                                                                                    m.p. 126—126.5°C
m.p. 162—163°C.
m.p. 162—163°C.
                                                                                                                                                                                                                                                                                         40
                                                                        II and II
                         Thus in the presence of Form II, Form I is converted to Form II at some temperature
                         Part 19-E: 7-O-Methyllincomycin hydrochloride (XLVII)
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A mixture of 3.08 gms. of 4-trans-propylhygric acid hydrochloride and 75 ccs. of acctonitrile was stirred magnetically in a 3-necked, 500 cc. flask, equipped with a drying tube and a thermometer extending below the liquid surface. On addition of 3.31 gms. of triethylamine, the solid dissolved rapidly to give a pale tan solution.

On cooling to -5°C. in an ice/methanol bath, a colorless precipitate of triethylammonium chloride separated. Without removal of the precipitate, 2.02 gms. (1.94 ccs.) of isobutyl chloroformate were added at such a rate that the temperature remained between -5°C. and 8°C., after which stirring was continued at -5°C. for 15 minutes. There were then rapidly added 2.0 gms. of methyl 7-O-methyl-1-thio-a-lincos-

aminide in 25 ccs. of water to the above mixed anhydride solution, giving a pale tan solution, which was stirred at 0°C. for 45 minutes. Thin-layer chromatography (silica gel, 8 ethyl acetate: 5 acetone: 1 water by volume) showed a trace only of residual aminosugar, and a major new zone of R = 0.4. Volatile solvent was removed in vacuo at 40°C, the tan aqueous residual solution adjusted to pH 10 by the addition of aqueous sodium hydroxide (N), the mixture extracted thrice with 100 cc. portions of chloroform, and the combined extracts washed with water and dried over anhydrous sodium sulfate. Removal of the solvent in vacuo at 40°C. give a tan amorphous solid (2.32 gms.).

Chromatography on silica gel (450 gms., column dimensions 3.8 × 95 cms.) in the system 1 methanol: 15 chloroform by volume following a forerum (250 ccs.) after which 25 cc. fractions were collected automatically, gave 7-O-methyllincomycin in fractions 44—70, inclusive, obtained on removal of the solvent in vactor as a colorless syrup (2.20 gms.). This syrup was dissolved in water (5 ccs.) by stirring and adding hydrochloric acid (concentrated) to attain a pH of 3, the solution filtered under suction, the sinter washed with water (3 ccs.) and the filtrate and washings cooled in an ice-methanol bath. With stirring, acetone (200 ccs.) was added, followed by ether (100 ccs.), giving a colorless crystalline precipitate which was collected and dried in a vacuum desiccator at room temperature. The solid (1.71 gms.) was obtained as small, elongated, colorless platelots, m.p. 155—157°C.

Anal. Calcd. for C₁, H₃₄O₄N₂S.HCl: C, 49.93; H, 8.16; N, 6.13; S, 7.02; Cl, 7.76; OMe, 6.79 M.W. (free base) 420.57. Found (corrected for 4.83% H₂O): C, 50.09; H, 8.22; N, 6.02; S, 7.20; Cl, 7.46; OMe, 7.03 [\$\alpha\$]₀ + 145°C (c, 1.063, H₂O) pKa' 7.6 Mol. Wt.: (mass spec., M⁺ of free base) 420

The 7(R)-O-methyllincomycin thus produced can be further processed by the novel process of this invention to yield the corresponding 3-nucleotide.

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Example 20

Part 20-A-1: Methyl N-acetyl-2-O-acetyl-3,4-O isopropylidene-1-thio-a-lincosaminide (XLVIII)

Methyl 6-N,7-O-ethylidyne-3,4-O-isopropylidene-1-thio-a-lincosaminide (5 gms.) (Example 1—C of U.S. Patent 3,337,527) was acetylated by leaving overnight a recomb temperature in a mixture of pyridine (25 ccs.) and acetic anhydride (12 ccs.). Removal of solvent on a rotating evaporator in pacuo at 40°C. gave a pale yellow syrup which was dissolved in chloroform, washed with water, saturated aqueous bicarbonate, again with water, and dried over anhydrous sodium sulfate. Thin-layer chromatography (silica gel, 75 methylethyl ketone: 25 acetone: 10 water by volume) showed the absence of starting material, and the formation of a new zone of slightly higher R. Removal of the solvent in vacuo at 40°C. gave methyl 2-O-acetyl-6N,7-O-ethylidyne-3,4-O-iso-propylidene-1-thio-a-lincosaminide as an almost colorless syrup which could not be induced to crystallize.

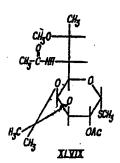
Water (75 ccs.) at pH 7 was added and, with magnetic stirring, the mixture was heated on a steam-bath. After 6 hours the solvent was removed in vacuo at 40°C, to give a colorless crystalline solid (5.95 gms.) which was chromatographed on slica (600 gms., column dimensions 4.8×79 cms.) in the system 1 methanol: 7 chloroform (by volume). After a 650 cc. forerun, 25 cc. fractions were collected autematically, the clution being followed by thin-layer chromatography. The desired material was present in fractions 35—41, inclusive. Removal of the solvent gave a colorless amorphous solid (1.57 gms.). Recrystallization from accome-Skellysolve B (technical hexane) gave colorless needles of methyl N-acetyl-2-O-acetyl-3,4-O-isopropylidene-1-thio-a-lincosaminide,

| Incomplete the control of the cont

lincosaminide (XLVIX)

Part 20-A-2: The procedure of Part 20—A—1 was repeated except that the solvent was removed a heating time of 2 hours (instead of 6 hours). The yield of methyl N-acetyl-2-O-acetyl-3,4-O-isopropylidene-1-thio-a-lincosaminide was increased to 60.5%.

Part 20—B: Methyl N-acetyl-2-O-acetyl-7-O-methyl-3,4-O-isopropylidene-1-thio-a-methyl-3,4-O-isopropylidene-1-



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Methyl N-acetyl-2-O-acetyl-3,4-O-isopropylidene-1-thio-a-lincosaminide (1.0 gm., 1 mol.), methyl iodide (37.6 gms., 16.5 ccs., 100 mols.), and silver oxide (3.1 gms., 5 mols.) were heated and stirred under gentle reflux for 16 hours. The methyl iodide was removed in vacuo at 40°C., and the resulting vellow-gray powder was extracted thoroughly with methylene chloride. Removal of the solvent in vacuo gave a yellow syrup (1.09 gms.). This crude product was subjected to countercurrent distribution (500 transfers) in the system 1 ethyl acetate: 1 ethanol: 1 water: 2 cyclohexane, by volume, using equal volumes of upper and lower phase. A major peak was found, of K=0.34, matching the theoretical curve.

Removal of the solvent from the combined fractions of the material of K=0.34 yielded a syrup (250 mgms.) which crystallized on standing. Recrystallization from ethyl

Removal of the solvent from the combined fractions of the material of K=0.34 yielded a syrup (250 mgms.) which crystallized on standing. Recrystallization from ethyl acetate-Skellysolve B, gave methyl N-acetyl-2-O-acetyl-7-O-methyl-3,4-O-isopropylidene-1-thio-or-lincosaminide as blunt, colorless needles, m.p. 152—154°C. (160 mgms.). A second recrystallization from the same solvent mixture yielded the pure product, m.p. 152.5—154°C.

Anal. Calcd. for C₁,H₂,O₇NS: C, 52.15; H, 7.47; N, 3.58; S, 8.18; N.W. 391.48. Found: C, 52.24; H, 7.48; N, 3.92; S, 7.98 Mol. Wt.: (mass spec., M⁺) 391 [α]₂ + 188- (c, 1.185, CHCl₃)

Part 20—C: Methyl N-acetyl-7-O-methyl-1-thio- α -lincosaminide and its triacetate

Methyl N-acetyl-2-O-acetyl-3,4-O-isopropylidene-1-thio-re-lincosaminide (100 mgms.) was stirred with water (20 ccs.) and aqueous hydrochloric acid (N, 5 ccs.) at room temperature overnight. The solution was neutralized by stirring with silver carbonate (3 gms.), the solids removed by filtration and washed with water, and the filtrate and washings taken to dryness on a rotating evaporator at 60°C./7 mm., giving methyl N-acetyl-7-O-methyl-1-thio-a-lincosaminide as a colorless syrup which did not crystallize. It was further characterized by converting it to the triacetate.

lize. It was further characterized by converting it to the triacetate.

Pyridine (5 ccs.) and acetic anhydride (3 ccs.) were added, the mixture swirled till the syrup had dissolved, and the mixture left overnight at room temperature. Solvent was then removed as completely as possible at 40°C./<1 mm., giving a tan crystalline mixture, which was dissolved in chloroform, washed with aqueous hydrochloric acid (N/10), water, saturated aqueous sodium bicarbonate, water, and dried over anhydrous sodium sulfate. Removal of the solvent in racuo gave methyl N-acetyl-2,3,4-tri-O-acetyl-7-O-methyl-1-thio-n-lincosaminide as an almost colorless crystalline solid which

acetyl-7-O-methyl-1-thio-n-lincosaminide as an almost colorless crystalline solid which separated from ethyl acetate-Skellysolve B in small colorless prisms, m.p. 211.5—213°C.

Anal. Calcd. for C₁,H₁,O₁NS: C₂ 49.64; H, 6.71; N, 3.22; S, 7.36; MeO, 7.13 N.W. 435.49. Found: C₂ 49.72; H, 6.77; N, 3.36; S, 7.27; MeO, 7.08 [a]_D + 229° (c₂ 0.7174, CHCl₂) Mol. Wt.: (mass spec., M⁺) 435

In the above Examples, in place of methyl iodide, there can be substituted ethyl, propyl, buryl, isobutyl, sec.buryl, and tert.buryl iodide to produce the corresponding 7-O-alkyl analogs.

Above in place of 4-propylhygric acid hydrochloride (1-methyl-4-trans-propyl-L-2-pyrrolidinecarboxylic acid hydrochloride) there can be substituted the hydrochlorides of other L-2-pyrrolidine carboxylic acids of the formula

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wherein R_2 is hydrogen or methyl or ethyl, and R_1 is hydrogen or $C_{1-\epsilon}$ alkyl, i.e., methyl, ethyl, propyl, butyl, pentyl, hexyl, heptyl, and octyl, and the isomeric forms thereof, to give compounds of the following formula

wherein R_2 and R_1 are as given above. The resulting lincomycin compounds can be converted to the corresponding novel 3-nucleotides by the novel processes of this invention.

WHAT WE CLAIM IS:-

1. A compound of the formula:

and the salts thereof, wherein Y can be in a- or 8-configuration and is -SR wherein R is alkyl of 1 to 6 carbon atoms, inclusive;

or —S—CH₂—CH₃—OH; R₁ is H is cis or trans-alkyl of from 1 to 8 carbon atoms; R₂ is H, CH₃ or C₂H₃; X is OH, chlorine, bromine, iodine or —OR₂, wherein R₃ is alkyl of 1 to 6 carbon atoms, inclusive, cycloalkyl, hydroxyalkyl or alkoxyalkyl each in the (R) or (S) configuration; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

2. The zwitterion form of the compound of claim 1.

3. A compound according to claim 1 hydroxyalkyl 15 20

3. A compound according to claim 1 having the formula:

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and the salts thereof, wherein Y, R, and R, are as given in claim 1, and Z is a nucleo-sude-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or

uridine.
4. A compound according to claim 3 having the formula:

$$\begin{array}{c|c} CH_3 & & & CH_3 \\ \hline & & & & CH_3 \\ \hline & & & & & CH_3 \\ \hline & & & & & & CH_3 \\ \hline & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & & \\ & \\ & & \\ & \\ & & \\ & \\ & & \\ & \\ & & \\ &$$

and the salts thereof, wherein Z is a nuceloside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

5. A compound according to claim 2 having the formula:

wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cyclidine or uridine and R_3 is CH_3 .

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6. A compound according to claim 1 having the formula:

and the salts thereof, wherein halo is chlorine or bromine and Y, R_1 , R_2 and Z are as given in claim 1.

7. A compound according to claim 6 having the formula:

and the salts thereof, wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine and R_0 is CH_2 .

8. A compound according to claim 2 having the formula:

wherein Z is a nucleoside-5' phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine and R_a is CH_a .

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9. A compound according to claim 1 having the formula:

and salts thereof, wherein halo is chlorine or bromine; R₃ is CH₃; R₁ is pentyl; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

10. A compound according to claim 9 having the formula:

and salts thereof, wherein R, is CH,; R, is pentyl; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

11. A compound according to claim 2 having the formula:

wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine; \hat{R}_3 is CH_3 ; and \hat{R}_1 is pentyl.

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12. A compound according to claim 1 having the formula:

and salts thereof, wherein halo is chlorine or bromine; R_2 is CH_3 ; R_1 is pentyl; R_2 is H; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

13. A compound according to claim 12 having the formula:

and salts thereof, wherein R, is CH₃; R₁ is pentyl; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

14. A compound according to claim 2 having the formula:

wherein R₂ is CH₃; R₁ is pentyl; and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

15. A compound according to claim 1 wherein Y is —SCH₂, R₁ is propyl, R₂ is CH₂, X is chlorine, and Z is a nucleoside-5'-phosphate group wherein said nucleoside is a nucleoside. is cytidine.

16. A compound according to claim 1 wherein Y is —SCH₃, R₁ is propyl, R₂ is CH₃, X is chlorine, and Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine.

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17. A compound according to claim 1 wherein Y is —SCH₂, R₁ is propyl, R₂ is CH₃, X is chlorine, and Z is a nucleoside-5'-phosphate group wherein said nucleoside is uridine.

18. A compound according to claim 1 wherein Y is —SCH₃, R₁ is propyl, R₂ is CH₃, X is chlorine, and Z is a nucleoside-5'-phosphate group wherein said nucleoside is groupseine.

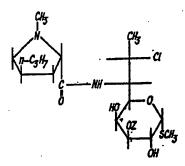
is guanosine.

19. A process for preparing a compound as defined in claim 1 which comprises incorporating a compound of the formula:

wherein R₁, R₂, X and Y are defined in claim 1, in the fermentation medium of a streptomyces fermentation.

20. A process according to claim 19 which comprises incorporating a compound of the formula:

in a Streptomyces coelicolor Müller, NRRL 3532, fermentation to produce compounds 15 of the formula:



wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cytidine or uridine.

21. A process according to claim 19 which comprises incorporating a compound of the formula:

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in a Streptomyces venezuelae, NRRL 3527 fermentation to produce a compound of the formula:

wherein Z is a nucleoside-5'-phosphate group wherein said nucleoside is adenosine, guanosine, cyrdiine or uridine.

22. A process for isolating a compound of the formula:

wherein R₁, R₂, Z, X and Y are as defined in claim 1, from a *Streptomyces* fermentation medium which comprises
(1) filtering the fermentation medium;
(2) absorbing the resulting filtrate on a suitable absorbent to remove water-soluble 10

- impurities,
- (3) chromatographing the resulting elaute from the absorbent on an anion exchange resin;
 - (4) subjecting fractions from the anion exchange resin to counter current distribution; and

(5) separating the individual 3-nucleotides by chromatography.
23. A therapeutic composition comprising, in unit desage form, from 25 to 500 mg. of a compound of the formula:

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wherein R₁, R₂, Z, X and Y are as defined in claim 1, or a pharmacologically acceptable salt thereof as an essential active ingredient in combination with a pharmaceutical carrier

24. A therapeutic composition comprising from 5% to 82% by weight of a compound of the formula:

wherein R₁, R₂, Z, X and Y are as defined in claim 1, or a pharmacologically acceptable salt thereof as an essential active ingredient in combination with a phrmaceutical vehicle.

25. A sterile composition for parenteral administration comprising from 5% to 82%, w/v, of a compound of the formula:

wherein R₁, R₂, Z₃ X and Y are as defined in claim 1, or a pharmacologically acceptable saft thereof as an essential active ingredient in combination with a sterile vehicle.

26. The process for treating susceptible microbial infectious disease in animals excluding humans which comprises administering to the bacterial host a therapeutic amount of a compound of the formula:

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wherein R₁, R₂, Z X and Y are as defined in claim 1, or a pharmacologically acceptable salt thereof in combination with a pharmaceutical carrier.

27. The process according to claim 26 for treating susceptible microbial infectious disease in animals excluding humans which comprises administering to the bacterial host, in unit dosage form, from 25 to 500 mg. of a compound of the formula:

wherein R₁, R₂, Z, X and Y are as defined in claim 1, or a pharmacologically acceptable salt thereof in combination with a pharmaceutical carrier.

28. A process according to claim 26 for treating susceptible microbial infectious disease in animals excluding humans which comprises administering to the infected host from 1 mg./kg. to 50 mg/kg. per day of a compound of the formula:

wherein R_1 , R_2 , Z, X and Y are as defined in claim 1, in combination with a pharmaceutical carrier.

29. A process of prophylactic treatment for the prevention of susceptible microbial infectious disease comprising the administering to a disease-susceptible animal host excluding humans an effective amount of a compound of the formula:

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wherein R1, R2, Z, X and Y are as defined in claim 1, or a pharmacologically acceptable salt thereof in combination with a pharmaceutical carrier.

30. A process according to claim 29 of prophylactic treatment for the prevention of susceptible microbial infectious disease comprising administering to a disease-susceptible animal host, excluding humans, in unit dosage form, from 25 to 500 mg. of a compound of the formula:

wherein R₁, R₂, Z₂, X and R are as defined in claim 1, or a pharmacologically acceptable salt thereof in combination with a pharmaceutical carrier.

31. A process for the preparation of a compound as claimed in any of claims 1 to 18 substantially as herein described with reference to Examples 1 to 10, 19 and

32. A compound as claimed in any of claims 1 to 18 when prepared by a process as claimed in claims 19 to 22.

33. A therapeutic composition comprising as the active ingredient a compound as claimed in any of claims 1 to 18 or 32 together with a pharmaceutically acceptable

34. A process for the treatment of microbial infectious disease in animals excluding humans which comprises administering to said humans and animals a compound as claimed in any of claims 1 to 18 or 32.

35. A process for the prevention of microbial infectious disease in animals excluding humans which comprises administering to the said animals a compound as claimed in any of claims 1 to 18 or 32.

36. A therapeutic composition comprising as the active ingredient a compound as claimed in claim 1 substantially as herein described with reference to Examples 11

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